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# (54) MAMMALIAN PROTEINS THAT BIND TO FKBP12 IN A RAPAMYCIN-DEPENDENT FASHION

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\*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

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### Related U.S. Application Data

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(51) Int. Cl.<sup>7</sup> ...... C12Q 1/68

(52) **U.S. Cl.** ...... **435/6**; 536/23.1; 536/23.4; 536/23.5; 536/24.3

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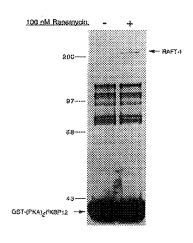
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57) ABSTRACT

A protein complex containing 245 kDa and 35 kDa components, designated RAFT1 and RAFT2 (for Rapamycin And FKBP12 Target) interacts with FKBP12 in a rapamycin-dependent manner. This interaction has the pharmacological characteristics expected from the observed in vivo effects of rapamycin: it occurs at low nanomolar concentrations of rapamycin and is competed by excess FK506. Sequences (330 amino acids total) of tryptic peptides derived from the affinity purified 245 kDa RAFT1 reveals striking homologies to the predicted products of the yeast TOR genes, which were originally identified by mutations that confer rapamycin resistance in yeast. A RAFT1 cDNA was obtained and found to encode a 289 kDa protein (2550 amino acids) that is 43% and 39% identical to TOR2 and TOR1, respectively.

#### 12 Claims, 10 Drawing Sheets



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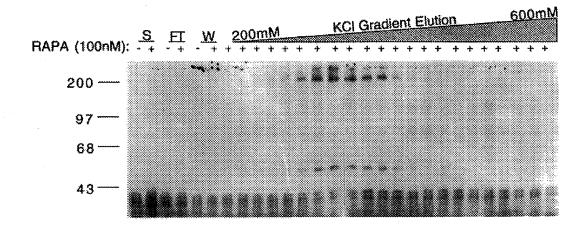
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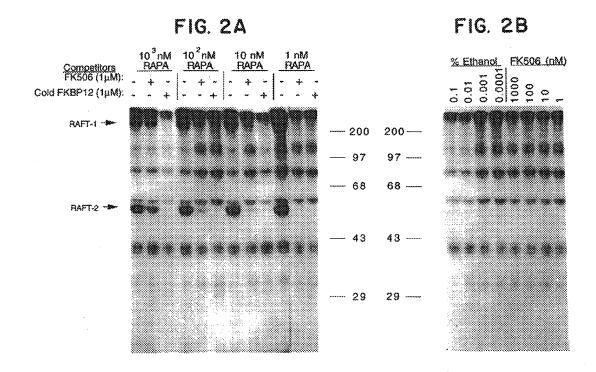
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FIG. 1



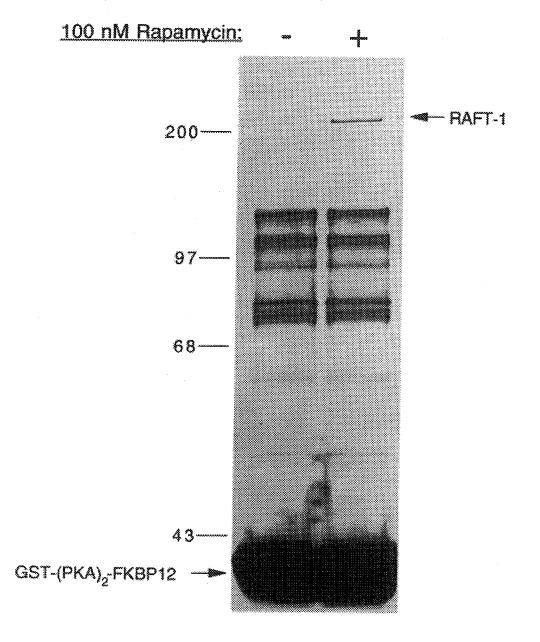
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MLGTGPATATAGAATSSNVSVLOOFASGLKSRNEETRAKAAKELOHYVIME SAGHIGKISFVDSELDTTFSTLNLIFDKLKSDVPOERASGANELSTTLISL TSSRFDGVVIGSNGDVNFKPILEKIFRELTSDYKEERKLASISLFDLLVSL	MEMASKAIGREAMAGDTFTAEYVEFEVK MRLAANTLGRETVPGGTLTSDFYEFEVR MRLAAKTLGKEAVPGGTYTSDFYEFEIK	LRACETLTTORERKEMOKPOWYRHTEEAEKGE EGKCETTIODROPALGKOWFORLFOGCTHGL LAKCESTLRNRDROLTSOWVORLATSCEYGF	DLMGFGTKPRHITPFTSFOAVOPOOSNALVGLLGYSSHOGLMGFGASPSPT 	医国DTOX CODT NAT SCVKKEKERTAAFOALGLETERYLKNIDMNAANNSDKPFILVSIGDETK-KXCHOIMDNYEILTNAPAKKIPHLKDDKPOILISIGD	GPG100D1-KELLEPMLAVGUSPALTAVLYDUSROTROUKKD10DGLUKME GPAFAKHLNKDLUNLMENCPMSDHMOETLM1ENEKTPSLESTVNSR1ENLU GRVLGKLLNRN1LDLMFKCPUSDYMOETFOILTERTPSLGPK1NDELLNLV	S DV ASETLALR <u>TAGS FEFEGHSUTOFVR</u> HCADHFLNS EHKE I RME AARTCS T DAOILIOCFKMOLIHHO-YSUTEFVRLITISYIEHEDSSVRK LAALTSC N DI KITIOKFRMUKNIKSR-FSLVEFVRIVALSYIEHTDPRVRK LAALTSC	LDERFOAHLAGAENLOALFVALNDOVFETRELATCTVGRLSSMNBAFYMPF LGSNFDPOLAGPDNLRLLFMALNDETFGTOLEATKTTGRLSSVNRAYVVPS UNPCRDPOLAGPDNLRLLFTALHDESFNTOSVAMELVGRLSSVNPAYVTPS	KDPDPDPNPGWINNV WATTGEWAONSGLEMRKWVDFWRVITMDMLODSSLL ODASSAWASTAWKVLGEWSVYGGKEMTRYLKEWMPLIINTFODOSNS ODTSSTWASTAWRTTGEWSVYGGEDMKIYLKDWFPLIIKTFODOSNS
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GLLGALDPYKHKVNIGMIDOSRDASAVSLSESKSSODSSDYSTSEMLVNNG	LNWIRVCDGAIREELFOOLGMLVSFVKSHIRPYMDEIVTLMREF	OLEGANIDDXLHILLEPINKLFDAPEVPLPSRKAALETVDRLTESLDF	INHORNDY #ICR VKGYT #ADEEEDPLI WOHRMLRSSOGD	SCWALLAOAYNPMARD <u>LFNAAFVSCWSEEN</u> EDOODELIRSIELALT	OKGPTPAILESUISIINNK LOOPEAASGVLEVAMKHFGELEIOATWYEK	AKMARMAAAAMGLGDWDSMEEYTCMIPRDTHDGAFYRAVLALHODLF	REIIROIWWERLOGCORIVEDWOKILHWRSLWVSPHEDMRTWLKYASL	- OHMOHEVOTMODOAOHAIATEDOOHKOELHK
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GILGAIDPYROKEREVTSTTDISTEONAPPIDIALLMOG	EDWMRTGSOSLLEEYFOOLCSLIIIVROHIRPHVDSIFOAIKDE	ESEGPNUEGXSHUITRKINOMAEFTSGNLORSAIITIGKUAKDVDL	IOHTI YBD#TNR LNNDV #PTKILEANTTDYKPAE-OMEAADAG-	ACSNEASMYYPLAKELFNTAFACVWTELYSOYOEDLIGSLCIALS	IKEPENSTIESUISINNOLNOTDAAIGILKHAOOH-HSLOLKETWFEK	KLIAPLAAGARWGLGEWDMLEOYISVMKPKSPDKEFFDAIEYEHKNDY	KLHYONLWTKRLLGCOKNWDLWORVERWRSLWIKPKODLOIWIKFANL	-NHLIGETSRLAHDLGLDPNNMIAOSVKLSSASTAPYVEEYTK
GLLGALDPY G1LGALDPY G1LGAIDPY	VMPTFLNW 11BG11LW 11PT1EDW	AA1OLEGAN KSLVTEGPN RLLESEGPN		PSERSOWAL ACERSOSSE HAERAOSNE	(O M M	ETOAK MARM EVKKAMAPL OTKKLIAPL	$\neg$ $\vdash$ $\bot$	IDAFOHMOH DEALKOLIN KEALNHLIG
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LBL-DEFYPAYSMVALMR SPSNDEYYLTYVIHNLMK SPSNDEYYTTVVIHCLK NTSIOSTLILLEOIVVA	1-1KLOIIILSVIESISKA V-AKLOITLVSVIEAISKA TDYASRIIIHPIVRTEDO SEMSSRIVOALVRIENNGD FEMSSRIVOALVRIENNGD	ALASGPVETGPMKKEHWST VTKEPWNOI	ODIAEVTOTLENLAEFHEH ENPPEIYOMLENLVEFHEH LNPPEIHOTLENLVEFHEH	LHEWEDALVAYDKKMDTNK LORWEDALAAYNEKEAAGE LERWEDALHAYNEREKAGD	SLWOOCIDKARDLEDAEET KKREVHIFNARDLEVTEES DNWSKHILNARDLEVTEIS	CGKSGRLALAHKTEVLLE- CRKSGRMALAKKVENTELE CRKSGRMRLANKALNMLEE	0 0 0 0 7 7 1 X X
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EAVLHYKHONOARDEKKKLRHASGANITNATTTATAASAAAATSTEGSNS EVISMLTSVSKKKOEGSDASSVTDIN-EFDNGHIGVNT EVISMVOEETKLNGGKNDDDDDDTAVNDNVRIDGSILGSGS	ERVETENEDYGHWRDVNERLVEGVKAIGIDTWEOXIPGEIARIDTRPLYG ERLETENET FGGIREATOAMHEGFNLIOIGTWEEYLPGEISRIHOPNOIXS ERLETELFNFGGIKEVSOAMYEGFNLMKIENWEEYLPGEISRIHOPDPTYS	AMMYSEELIRVAI EWHEMWHEGLEEASRLYFGERNVKGMFEVLEPLHAMME AELNSHELIRMAV EWHEOWYEGLDDASROFFGEHNTEKMFAALEPLYEMLK AELYSHELIRVAV EWHELWYEGLEDASROFFVEHNIEKMFSTLEPLHKHLG	10 % %	KNESTORYAVTPUSTNSGUIGWYRHCDTLHAUTRDYREKKKTLUNTEHRIN RHIDIOOYPAIRUSPKSGULGWYRNSDTFHVUTREHREAKKIPUNTEHWVN RHUDIOOYPAIRUSPKSGULGWYRNSDTFHVUTREHRDAKKIPUNTEOWVY	SLAVMSMVGYILGLGDRHPSNLMLDRLSGKILHIDFGDCFEVAMTREKFPE SLAVMSMTGYILGLGDRHPSNLMLDRITGKVIHIDFGDCFEAAILREKFPE SLAVMSMTGYILGLGDRHPSNLMLDRITGKVIHIDFGDCFEAAILREKYPE	NWRLMDTNAKGNKRSRTRTDSYSAGOSVEILDGVELGERAHKKIGTTV NWGFDLBTKKIEEETGIOL HWGFDLPPOKLTEOTGIPL	DTILDVPTOVELLIKOATSHENECOCYIGWCPFW NDLDVPEOVDKLIOOATSVENECOHYIGWCPFW NEUDVPEOVDKLIOOATSIERLCOHYIGWCPFW
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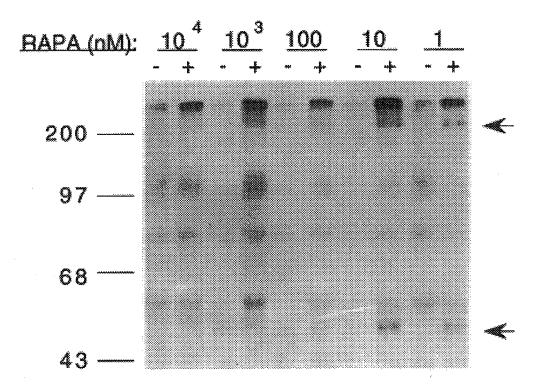
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FIG. 5



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# MAMMALIAN PROTEINS THAT BIND TO FKBP12 IN A RAPAMYCIN-DEPENDENT FASHION

This application is a continuation-in-part of application <sup>5</sup> Ser. No. 08/265,967, filed on Jun. 27, 1994.

This invention was made with government support under MH18501, DA00266, and DA00074, awarded by the National Institutes of Health. The government has certain rights in this invention.

#### BACKGROUND OF THE INVENTION

The natural products cyclosporin A, FK506, and rapamycin are potent immunosuppressants with realized or potential clinical applications in the prevention of graft rejection after organ transplantation and the treatment of autoimmune disorders (Borel, 1986; Kino et al., 1987; Martel et al., 1977). These drugs act by inhibiting intermediate steps in the signaling pathways that effect the T-cell response to antigen (for reviews see, Fruman et al., 1994; Kunz and Hall, 1993; Liu, 1993; Schreiber, 1991). This makes them useful probes for identifying the components of those pathways and determining their physiological roles.

The immunosuppressants interact with the 25 immunophilins, which are small, soluble, receptor proteins that mediate their actions. Cyclosporin A (a cyclical undecapeptide) binds to cyclophilin A, whereas FK506 and rapamycin (two related macrolide antibiotics) bind to a distinct receptor protein, FKBP12 (Handschumacher et al., 30 1984; Harding et al., 1989; Siekierka et al., 1989). Though cyclophilin and FKBP12 differ markedly in amino acid sequence, both immunophilins have peptidyl-prolyl cistrans isomerization (rotamase) activity, which is inhibited by their respective ligands (for review, see Heitman et al., 35 1992). However, this inhibition does not appear to explain the effects of the immunosuppressants (Bierer et al., 1990a, b; Tropschug et al., 1989). Instead, the action of cyclosporin A and FK506 derives from the binding of the drug-receptor complexes to the calcium-activated protein phosphatase, calcineurin (Liu et al., 1991). This association inhibits the catalytic activity of the phosphatase, which is required for the Ca++-dependent initial step in the activation of the T-lymphocyte via the T-cell receptor (Flanagan et al., 1991; Kronke et al., 1984).

On the other hand, rapamycin appears to block a later, Ca++-independent stage in the T-cell response. This drug selectively inhibits the IL-2 stimulated G1 to S cell-cycle transition that initiates T-cell proliferation (Dumont et al., 1990b). Although this inhibition has been correlated with the 50 decreased activity of the 70 kDa S6 kinase (pp70<sup>S6K</sup>), a known downstream effector of the IL-2 receptor, the FKBP12-rapamycin complex does not appear to interact directly with this kinase (Chung et al., 1992; Kuo et al., 1992). Similarly, in T-cells and other cell types, rapamycin 55 blocks progression of the cell cycle by preventing the activation of the cyclin-dependent kinases p33cdk. p34cdc2, but an association of the drug-immunophilin complex with the kinases or their respective cyclins has not been demonstrated (Albers et al., 1993; Jayaraman and Marks, 60 1993; Morice et al., 1993).

In the budding yeast *S. cerevisiae*, rapamycin also causes an arrest in the G1 phase of the cell cycle through its binding to a highly conserved FKBP12 homologue (Heitman et al., 1991b). The toxicity of the drug for yeast cells has allowed, 65 through genetic selection, the identification of two homologous genes, which, when mutated, render the cells

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rapamycin-resistant (Heitman et al., 1991a). This led to the proposal that the products of these genes, which show some amino acid homology to the catalytic domain of the p110 subunit of PI-3 kinase, are the Targets Of Rapamycin and 5 hence to the designation of the genes as TOR1 and TOR2 (Kunz et al., 1993). Direct support for this proposal, however, has not been presented and how the TOR gene products confer sensitivity to rapamycin remains to be elucidated. Alternatively, it has been suggested that in the signaling pathway blocked by rapamycin, the TOR proteins, like the S6 kinase and the cyclin-dependent kinases, lie downstream from the direct target of the FKBP12-rapamycin complex (Albers et al., 1993; Helliwell et al., 1994). This model assumes that the TOR mutations lead to the constitutive activation of the TOR1 and TOR2 proteins.

Besides binding to calcineurin in a FK506-dependent manner, FKBP12 can also interact with calcium-channel proteins, the ryanodine receptor, which mediates calcium induced calcium released (Jayaraman et al., 1992; Timerman et al., 1993) and the inositol 1,4,5,-triphosphate (IP<sub>3</sub>) receptor (A. Cameron, A. Kaplin, D. Sabatini, J. Steiner, S. Snyder, unpublished). These associations do not require FK506 or rapamycin; indeed these drugs dissociate the FKBP12-channel complex.

There is a need in the art to identify, isolate, and purify the mammalian cellular proteins that interact with FKBP12 only in the presence of rapamycin. Such proteins play a role in immunological, neurological, and cell cycle functions.

#### SUMMARY OF THE INVENTION

It is an object of the invention to provide isolated, purified cDNA molecules encoding rapamycin and FKBP target molecules.

It is another object of the invention to provide fusion proteins comprising rapamycin and FKBP targets.

It is still another object of the invention to provide an isolated and purified rapamycin and FKBP target molecule.

It is still another object of the invention to provide an expression construct which directs synthesis in a cell of an RNA molecule which inibits expression of a rapamycin and FKBP target molecule.

It is yet another object of the invention to provide isolated, purified cDNA molecules which are complementary to genes encoding rapamcyin and FKBP target molecules.

It is an object of the invention to provide a method of screening for potential therapeutic agents.

It is another object of the invention to provide a method of purifying a rapamycin and FKBP target molecule.

It is still another object of the invention to provide a method of isolating DNA sequences which code for rapamycin and FKBP target molecules.

These and other objects of the invention are provided by one or more of the embodiments described below. In one embodiment of the invention an isolated, purified cDNA molecule is provided which encodes RAFT1, a protein having the amino acid sequence shown in SEQ ID NO:2.

In another embodiment of the invention a fusion protein comprising the amino acid sequence shown in SEQ ID NO:2 is provided.

In yet another embodiment of the invention an isolated and purified RAFT1 protein having the amino acid sequence shown in SEQ ID NO:2 is provided. Also provided is an isolated and purified RAFT2 protein, having an apparent molecular weight on SDS polyacrylamide gels of 35 kDa. Also provided is an isolated and purified mammalian RAFT

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protein which is free of proteins which do not bind to rapamycin and FKBP12. Also provided is a mammalian RAFT protein prepared by the process of:

contacting a preparation of mammalian proteins with FKBP12 in the presence of rapamycin;

isolating mammalian proteins which bind to FKBP12 in the presence of rapamycin from those mammalian proteins which do not bind; and

dissociating bound mammalian proteins from FKBP12 to provide a mammalian RAFT protein.

In still another embodiment of the invention an expression construct is provided. The expression construct comprises a promoter operably linked to at least 20 nucleotides of the antisense strand of RAFT1 cDNA, said expression construct directing synthesis in a cell of an RNA molecule which is complementary to RAFT1 RNA.

In another embodiment of the invention an isolated, purified cDNA molecule comprising at least 20 nucleotides of the sequence as shown in SEQ ID NO:1 is provided.

In yet another embodiment of the invention a method of screening substances for potential as therapeutic agents is provided. The method comprises the steps of:

contacting a substance to be tested with three components: (a) FKBP12, (b) rapamycin, and (c) a protein 25 selected from the group consisting of RAFT1 and RAFT2;

determining the amount of one of said components bound to the other components in the presence and absence of said substance; a substance which increases or 30 decreases the amount of said component bound being a potential therapeutic agent.

In one embodiment of the invention a method of purifying a mammalian RAFT protein is provided. The method comprises the steps of:

contacting a preparation of mammalian proteins with FKBP12 in the presence of rapamycin;

isolating mammalian proteins which bind to FKBP12 in the presence of rapamycin from those mammalian proteins which do not bind;

dissociating bound mammalian proteins from FKBP12 to provide a mammalian RAFT protein.

In another embodiment of the invention methods of isolating mammalian RAFT DNA sequences are provided.

One of the methods comprises:

probing a library of mammalian DNA sequences with a probe which comprises at least 15 contiguous nucleotides selected from the sequence shown in SEQ ID NO:1.

Another of the methods comprises:

amplifying a DNA sequence using at least one primer which comprises at least 10 contiguous nucleotides selected from the sequence shown in SEQ ID NO:1.

These and other embodiments of the invention provide the 55 art with potent tools for identifying drugs useful in the treatment of immunological, neurological, and cell cyclerelated diseases and defects.

#### BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows partial purification of the FKBP12-rapamycin target proteins from brain cytosol by heparin column chromatography. A cytosolic fraction prepared from a rat brain homogenate was applied to a heparin column. The material that remained bound to the column after washing 65 with 5 column volumes of wash buffer containing 200 mM KCl, was eluted with a linear gradient from 200 mM to 600

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mM KCl in homogenization buffer. Aliquots of the crude cytosol (S), the column flow through (FT), and the wash (W) were tested in the crosslinking assay with (+) or without (-) rapamycin (100 nM). Every other fraction eluted from the heparin column was tested in the crosslinking assay in the presence of 100 nM rapamycin. No rapamycin specific crosslinked products are visible in the crude cytosol because of the high concentrations of endogenous FKBP12 present in the initial sample.

FIG. 2 shows FK506 and unlabeled FKBP12 prevent the rapamycin-dependent association of <sup>32</sup>P-FKBP12 to the target proteins.

FIG. 2A) The heparin column eluate containing the RAFTs was tested in the crosslinking assay at the indicated concentrations of rapamycin with or without the addition of 1  $\mu$ M FK506 or 1  $\mu$ M FKBP12. FIG. 2B) Neither FK506 alone nor the ethanol vehicle induce crosslinking of FKBP12 to RAFT. The heparin eluate containing RAFT was tested in the crosslinking assay with the indicated concentrations of FK506 or ethanol. This experiment was repeated twice with identical results.

FIG. 3 shows purification of RAFT1 with a FKBP12-rapamycin affinity column. RAFT enriched fractions eluting from the heparin column between 300 and 450 mM KCl, were incubated in the presence (+) or absence (-) of 100 nM rapamycin with GST-(PKA)2-FKBP12 fusion protein (20 μg) immobilized on glutatlione agarose beads. The material that remained associated with the beads after extensive washes was analyzed by SDS-PAGE (8%) and silver staining. RAFT1 is present only in the sample treated with rapamycin. The large band at 36 kDa is the GST-FKBP12 fusion protein.

FIG. 4 shows the alignment of RAFT1 amino acid sequence with the predicted amino acid sequences of TOR2 (SEQ ID NO:4) and TOR1 (SEQ ID NO:3).

The alignment was maximized by introducing insertions marked by dashes. Sequences in RAFT1 identical to TOR2 and/or TOR1 are indicated with gray shading. The sequences of tryptic peptides obtained by microsequencing are indicated with a line above the RAFT1 sequence. Sequences used to design primers for PCR are indicated with an arrow above the residues (direction indicate sense or antisense). The PKC site conserved between RAFT1, TOR1 and TOR2 is boxed.

FIG. 5 shows rapamycin-dependent crosslinking of FKBP12 to two PC12 cell cytosolic proteins of approximate molecular weight 245 kDa and 35 kDa.

<sup>32</sup>P-labeled FKBP12 (10<sup>5</sup> cpm) was incubated with cytosolic fractions from PC12 cells with or without the indicated concentration of rapamycin for 1 hr. at 4° C. The crosslinker DSS was then added and the incubation continued for 40 minutes before processing for SDS-PAGE (4%–12% gradient) and autoradiography. The arrows indicate the two Lands that appear only in the presence of rapamycin. This experiment was repeated three times with identical results.

# DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

We have isolated and identified proteins, which we designate RAFT1 and RAFT2, that interact with the FKBP12-rapamycin complex. Rapamycin-induced binding of FKBP12 to RAFT1 occurs at drug concentrations as low as 1 and 10 nM, resembling pharmacological potency in vivo (Bierer et al., 1990a; Dumont et al., 1990a). FK506 and 65 rapamycin bind with similar affinities to the same binding site on FKBP12 and antagonize each others' actions in vivo (Bierer et al., 1990a; Dumont et al., 1990b). Consistent with

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these facts, FK506 does not induce interactions between FKBP12 and RAFT1 but, instead, prevents the rapamycin-mediated effect. Since rapamycin has pleiotropic effects on a wide variety of cell types, the target of its complex with FKBP12 is likely to be an early participant in several signal transduction pathways.

We have also isolated and purified a cDNA molecule which encodes RAFT1. The nucleotide sequence of RAFT1 is shown in SEQ ID NO:1. The predicted amino acid sequence of the protein, which exactly corresponds to the 10 empirically determined amino acid sequences of tryptic peptides of RAFT1, is shown in SEQ ID NO:2. The cDNA sequence can be used to express in recombinant cells RAFT1 proteins or portions of the RAFT1 protein. Similarly, the cDNA sequence can be used to construct fused genes which will express fusion proteins comprising all or part of the RAFT1 sequence. Having provided the art with the amino acid sequence of the RAFT1 protein, other coding sequences can be devised which differ from that shown in SEQ ID NO:1 by virtue of the degeneracy of the genetic 20 code. Such nucleotide sequences are within the scope of the present invention.

RAFT1 has an apparent molecular weight on SDS polyacrylamide gels of 245 kDa. RAFT2 has an apparent molecular weight on SDS polyacrylamide gels of 35 kDa. 25 Isolated and purified RAFT1 protein can be obtained by means of recombinant DNA technology or by isolating and purifying the protein directly from natural sources. One means of purifying RAFTs involves contacting a preparation of mammalian proteins with FKBP12 in the presence of 30 rapamycin. Those proteins which bind to FKBP12 in the presence of rapamycin can then be separated from those which do not bind. Bound proteins can then be dissociated to yield a preparation of a RAFT protein. It is convenient if the FKBP12 is immobilized, for example, on a solid support. 35 One convenient means is to immobilize FKBP12 on a column-packing matrix. For example, an FKPB12glutathione-S-transferase fusion protein can be readily bound to glutathione-agarose to provide immobilized FKBP12. Another means of purifying RAFT proteins is by 40 use of a heparin chromatography column. The RAFT proteins bind to the heparin and can be eluted at 300 to 450 mM

Because of the role of rapamycin in immunological, cell cycle, and neurological functions, it may be desirable to 45 inhibit the expression of RAFT1. One means to accomplish this is to use antisense polynucleotides. Antisense polynucleotides can be made synthetically, according to the sequence provided in SEQ ID NO:1. Alternatively, expression constructs may be used which comprise a promoter operably linked to at least 20 nucloetides of the antisense strand of RAFT1 cDNA. The expression construct directs the synthesis in a cell of an RNA molecule which is complementary to RAFT1 mRNA. Any suitable promoter can be used, depending on the cell system in which expression of the antisense molecule is desired.

The nucleotide sequence of SEQ ID NO:1 can be used to generate probes which comprise at least 15–20 nucleotides of the recited sequence. In some cases probes of 25, 30, 35, 40, 50, or 100 nucleotides may be desired. These probes can 60 be used to screen a library of mammalian DNA molecules. Techniques for making nucleotide probes and screening genomic or cDNA libraries are well known in the art. Alternatively, other RAFT nucleotide sequences can be obtained by amplification of mammalian DNA using as 65 primers one or two polynucleotides comprising at least 10 contiguous nucleotides selected from the sequence shown in

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SEQ ID NO:1. Techniques for amplification of DNA are also well known in the art.

RAFT1 and RAFT2 can be used to screen substances for potential as therapeutic agents for immunological, cell cycle, and neurological disease states. As described here, rapamycin, FKBP12, RAFT1, and RAFT2 bind to each other and form a complex. Test compounds can be screened for potential therapeutic utility by contacting a test compound with three components: (a) FKBP12; (b) rapamycin; and (c) a protein selected from the group consisting of RAFT1 and RAFT2. The amount of one of the components in the complex is determined, in the presence and in the absence of the substance to be tested. A substance which increases or decreasees the amount of the component in the comlex is a potential therapeutic agent. Means used for determining amounts of components can be any known in the art, including the use of radioactive components, antibodies specific for components, densitometry, etc.

#### **EXAMPLES**

The following materials were used in the examples described below. Frozen rat brains stripped of the meninges were obtained from Harlan Bioproducts (Indianapolis, Ind.). Other materials were purchased from the following sources:  $[\gamma^{-32}P]$ -ATP (NEG-02z) from New England Nuclear (Cambridge, Mass.), glutathione-agarose, heart muscle kinase (PKA, #P2645), and heparin-agarose from Sigma Chemical (St. Louis, Mo.), thrombin and antithrombin from Boehringer Mannheim (Indianapolis, Ind.), and disuccinimidyl suberate (DSS) from Pierce (Rockford, Ill.). Rapamycin was a gift of the Wyeth-Ayerst company (Philadelphia, Pa.) and FK506 a gift of the Fujisawa company (Tsukuba City, Japan).

#### Example 1

Rapamycin Promotes the Binding of FKBP12 to Two Cytosolic Proteins of Mr 245 and 35 kDa

A <sup>32</sup>P-radiolabeled FKBP12 probe was used to detect proteins that associate with the immunophilin in the presence of ligand, and are crosslinked to it by the bivalent reagent DSS. The probe was prepared by phosphorylating with [γ<sup>32</sup>P]ATP a recombinant rat FKBP12 to which two consensus sites for cyclic AMP-dependent protein kinase (PKA) were added at the N-terminus (Blanar and Rutter, 1992; Li et al., 1992). Since this modification did not alter the capacity of the protein to associate with calcineurin in the presence of FK506, the probe can be used to identify a target of the FKBP12-rapamycin complex.

PC12 pheochromocytoma cell cytosolic extracts were incubated with 32P-FKBP12 in the presence or absence of rapamycin and then treated with the crosslinker DSS before gel electrophoretic analysis followed by autoradiography. The drug caused the formation of two protein complexes with radioactive FKBP12, corresponding to bands of Mr 260 and 50 kDa (FIG. 5). Taking into account the 15 kDa Mr of the modified FKBP12 probe, the crosslinked proteins were estimated to be 245 kDa and 35 kDa, respectively. The crosslinked complexes were observed over a wide rapamycin concentration range, but were more prominent at the low concentrations of 1 and 10 nM, possibly because of an inhibitory effect on the association of the higher amounts of ethanol (the solvent of the drug) present at the higher drug concentrations (FIG. 5). Rapamycin also induced the formation of similar complexes when the probe was incubated with cytosolic extracts from several rat tissues, including

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liver, kidney, heart, small intestine, thymus, testes, spleen and brain, but no significant differences in abundance of the crosslinked proteins between the tissues were observed. For convenience, further experiments were carried out with whole brain extracts.

The formation of the rapamycin-dependent complexes was specific for FKBP12, since in similar experiments with the related immunophilin <sup>32</sup>P-FKBP25, no ligand induced complexes were observed.

PC12 cells were maintained in culture as described (Altin et al., 1991). PC12 cells were lysed in homogenization buffer with 0.3% NP-40 instead of CHAPS. Lysis was accomplished in 2 ml buffer/T-150 flask by repeated vortexing at 4° C. Cell debris was sedimented by centrifugation for 10,000×g for 10 minutes at 4° C.

The labeled, cleaved FKBP12 was diluted to 10,000 cpm/ml in 50 mM Hepes pH 7.5, 1 mg/ml BSA. 10  $\mu$ l of labeled protein (100,000 cpm total), 10  $\mu$ l of tissue or PC12 cell extract, and 10  $\mu$ l of drug dilutant buffer (20 mM Hepes 6.8, 100 mM KCl, 1 mM EGTA, 1 mM DTT) containing either 3-fold the desired final concentration of rapamycin, FK506, or equivalent amounts of ethanol, were mixed and incubated for 1 hour at 4° C. After this incubation, 1 ml of 5.5 mg/ml disuccinimidyl suberate (DSS) was added and the incubation continued for 40 minutes. The reaction was terminated by adding one column volume of 2× concentrated sample buffer (Laemmlli, 1970) containing 50 mM Tris pH 7.4 and processed by SDS-PAGE (10%, unless otherwise specified) and autoradiography.

#### Example 2

Specificity of the Rapamycin Induced Association: the Interaction of <sup>32</sup>P-FKBP12-Rapamycin with the 245 and 35 kDa Proteins is Competed by FK506 and by Unlabeled FKBP12

To investigate further the specificity of the interaction of <sup>32</sup>P-FKBP12-rapamycin with the cytosolic proteins, we performed a partial purification to remove endogenous FKBP12, which is present in brain at high concentrations (Steiner et al., 1992). This was accomplished by chromatography on a heparin column, to which the cytosolic proteins that interact with FKBP12-rapamycin bound and could be eluted at 300 to 450 mM KCl (FIG. 1). Free FKBP12, on the other hand, was recovered in the flow-through of this column, as demonstrated by binding to [<sup>3</sup>H]FK506 (data not shown).

The rat brain extract was applied to a heparin column (2 ml of packed heparin-agarose per brain) at a flow rate of 1.5 ml/min. The column was washed with 10 column volumes of buffer (20 mM Hepes pH 6.8, 200 mM KCl, 1 mM EGTA, 50 mM NaF, 1.5 mM Na $_3$ VO $_4$ , 4 mM DTT, 1 mM PMSF) and the same protease inhibitors as in the homogenization buffer. The material bound to the column was eluted with a linear KCl gradient from 200 to 600 mM in homogenization buffer. Aliquots (10  $\mu$ l) of the fractions collected were tested in the crosslinking assay and positive fractions were pooled and concentrated in a centriprep-100 (Amicon, Beverly, Mass.) to  $\frac{1}{2}$  starting volume. The flowthrough of the heparin column was assayed for the presence of FKBP with a  $^3$ H-FK506 binding assay, as described (Steiner et al, 1992).

FK506 antagonizes actions of rapamycin, and both drugs compete for the same binding site on FKBP12 (Bierer et al., 1990a; Dumont et al., 1990a). Accordingly, we examined the 65 influence of FK506 on the rapamycin-induced interaction of <sup>32</sup>P-FKBP12 with its putative cytosolic targets. At concen-

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trations ranging from 1 nM to 1  $\mu$ M rapamycin induced the appearance of intense bands representing crosslinked proteins and, at all rapamycin concentrations tested, this effect was antagonized by 1  $\mu$ M FK506 (FIG. 2A). As expected for ligands of similar affinity for FKBP12, when equal concentrations (1  $\mu$ M) of rapamycin and FK506 were present, the intensities of the crosslinked bands were reduced by approximately 50% and the reduction progressively increased with increasing ratios of FK506/rapamycin. The heparin column eluate apparently contains limiting amounts of the putative targets of the FKBP12-rapamycin complex, since excess unlabeled FKBP12 (1  $\mu$ M) completely suppressed the appearance of the crosslinked bands containing labeled FKBP12 (FIG. 2A).

Control experiments (FIG. 2B) confirmed the specificity of the rapamycin effect since the formation of the complex was not induced by several concentrations of FK506 or by ethanol, the solvent of the drugs. These experiments demonstrate that the crosslinked proteins are specific targets of the FKBP12-rapamycin complex and not of the FKBP12-FK506 complex, nor of FKBP12 alone. Therefore, we designate the crosslinked proteins RAFT1 (245 kDa) and RAFT2 (35 kDa) for Rapamycin And FKBP12 Target.

We attempted to separate RAFT1 and RAFT2 under nondenaturing conditions by several chromatography and gel filtration procedures, including DEAE and CM cellulose, reactive dye green 5, and Superose 6 (data not shown). All of these efforts failed, suggesting that RAFT1 and RAFT2 are part of a complex, although it is possible that RAFT2 is a proteolytic fragment of RAFT1 that contains the FKBP12-rapamycin binding site and remains tightly bound to the rest of the polypeptide.

#### Example 3

#### Purification of RAFT1

We purified RAFT1 from the heparin column eluate based on its affinity for FKBP12-rapamycin. We constructed a glutathione-S-transferase-FKBP12 fusion protein by cloning, in frame downstream of GST, a cDNA encoding FKBP12 with two N-terminal PKA consensus sites (Smith and Johnson, 1988; Blanar and Rutter, 1992; Li et al., 1992). The encoded protein was expressed in bacteria, purified and immobilized on glutathione-agarose beads. SDS-PAGE analysis of the beads recovered after incubating them with the heparin eluate in the presence or absence of rapamycin shows that the drug induces the binding to the beads of a protein of 245 kDa (FIG. 3). With this simple purification scheme we were able to purify about 5  $\mu$ g of RAFT1. A low transfer efficiency to nitrocellulose membrane resulted in only 2.5  $\mu$ g being available for protein sequencing, which corresponds to 10 picomoles of a protein of this size.

Standard techniques of molecular biology cloning were used as described (Sambrook et al., 1989) for the preparation of GST-(PKA)<sub>2</sub>-FKBP12 and GST-(PKA)<sub>2</sub>-FKBP25 fusion proteins, unless otherwise specified. All cDNAs obtained with the polymerase chain reaction were sequenced using the Sequenase kit (Amersham, Arlington Heights, Ill.). cDNAs for the rat FKBP12 and FKBP25 were obtained with the PCR using 5' and 3' primers to the corresponding human FKBP12 (Standaert et al., 1990) or FKBP25 (Jin et al., 1992) sequences. The cDNAs were cloned into pBluescript (Stratagene, La Jolla, Calif.).

A 5' primer (PKA-12-1 or PKA-25-1) encoding a BamHI site, two consensus PKA phosphorylation sites (Blanar and Rutter, 1992; Li et al., 1992), and the first 6 amino acids of

FKBP12 or FKBP25 was used with a 3' primer (PKA-12-2 or PKA-25-2) encoding an EcoRI site and the last 6 codons of FKBP12 or FKBP25 in a PCR with Vent Polymerase (New England Biolabs, Beverly, Mass.) using the rat FKBP cDNAs cloned in pBluescript as templates. The amplified DNA fragments were gel purified, digested with BamH1 and EcoR1 and cloned into the pGEX-2T vector (Pharmacia, Upsala, Sweden) that had been linearized with the same restriction enzymes. The resulting construct was used to transform BL21 (DE3) E. coli (Novagen, Madison, Wis.) in 10 which expression can be induced with IPTG.

The primer sequences were as follows:

PKA-12-1: 5' CCGGATCCCGTCGAGCTTCAGT-TGAACTACGGCGTGC TTCTGTAGCCATGG-GAGTGCAGGTGGA 3' (SEQ ID NO:5)

PKA-12-2: 5' GGCCGGAATTCTCATTCCAGTTTTA-GAA 3' (SEQ ID NO:6)

PKA-25-1: 5' CCGGATCCCGTCGAGCTTCAGT-TGAACTACGGCGTGC TTCTGTAGCCATGGCG-GCGGCCGTTCC 3' (SEQ ID NO:11)

PKA-25-2: 5' GGCCGGAATTCTCÁATCAATATC- 20 CACTA 3' (SEQ ID NO:12)

The fusion proteins were purified with glutathione-agarose as previously described (Smith and Johnson, 1988) from bacterial cultures induced with 1 mM IPTG.

The concentrated heparin column eluate was incubated 25 for 2 hours at 4° C. with ½0 volume of glutathione-agarose to remove endogenous glutathione binding proteins. The beads were removed by centrifugation at 1000xg for 3 minutes. Fresh glutathione-agarose ( $\frac{1}{500}$  volume) and 20  $\mu$ g of purified GST-PKA-FKBP12 fusion protein were then 30 added to the cleared heparin column eluate with or without 100 nM rapamycin. After a 1 hour incubation at 4° C., the bead were washed 5x with 1.5 ml ice-cold PBS containing 1% Triton X-100 and 500 mM NaCl. The beads were transferred to 3x volume SDS-PAGE sample buffer, and the eluted proteins fractionated by SDS-PAGE and the gel silver stained

Whether RAFT2 was also bound to the beads could not be determined in this experiment, because its presence would be masked by the large band of similar Mr corresponding to the GST-(PKA)<sub>2</sub>-FKBP12 fusion protein. When smaller fusion proteins, such as an epitope-tagged FKBP12, were employed for the affinity matrix, the binding of the 35 kDa RAFT2 could also be observed.

The immunophilin fusion proteins containing N-terminal phosphorylation sites for PKA were labeled with a modifi- 45 cation of published procedures (Blanar et. al., 1992, Li et. al., 1992). 10 ng of purified GST-PKA-FKBP12 or 25 was mixed with 40 units of PKA and 100 mCi of [y-P<sup>32</sup>]-ATP in a buffer containing 20 mM Hepes pH 7.7, 100 mM NaCl, 12 mM MgCl<sub>2</sub>, 1 mM DTT.

After a 1.5 hour at 37° C. the incubation mixture containing labeled fusion protein was dialyzed twice against 1L of thrombin cleavage buffer (50 mM Tris pH 7.4, 150 mM NaCl, 2.5 mM CaCl<sub>2</sub>). The labeled fusion protein was buffer containing 2 mg/ml thrombin and incubating at room temperature for 2 hours. The thrombin was inactivated by adding an equal volume of a stop solution consisting of 1 mM DTT, 1 mM PMSF, 100 units/ml antithrombin III. The specific activity of the probes was estimated at 1×10<sup>5</sup> cpm/pmol of the protein.

#### Example 4

Protein Sequencing of RAFT1: Homology to TOR1 and TOR2

Affinity purified RAFT1 was separated by SDSpolyacrylamide gel electrophoresis from other proteins that 10

adsorbed to the glutathione-agarose beads, transferred to nitrocellulose membrane, and digested with trypsin. Fractionation of the tryptic digest by narrow-bore reverse phase chromatography yielded a complex pattern of over a hundred peaks whose purity was assessed by mass spectroscopy. In most cases, the peaks exhibited multiple mass to charge peak values and it was necessary to rechromatograph these peak fractions on a microbore columns of different selec-

For protein sequence analysis affinity purified material derived from 50 brains was fractionated by SDS-PAGE and transferred to nitrocellulose membranes. The proteins transferred were visualized by Ponceau S staining, the 245 kDa RAFT1 band excised and processed for internal amino acid sequence analysis, essentially as described (Tempst et al., 1990).

Membrane-bound protein, about 2.5  $\mu$ g, was subjected to in-situ proteolytic cleavage using 1 µg trypsin (Sequencing Grade; Boehringer-Mannheim) in 25 ml 100 mM NH<sub>4</sub>HCO<sub>3</sub> (supplemented with 10% acetonitrile and 3% Tween-80) at 37° C. for 3 hours. The resulting peptide mixture was reduced and S-alkylated with, respectively, 0.1% β-mercapto ethanol and 0.3% 4-vinyl pyridine, and fractionated by two-dimensional reversed phase HPLC.

For the primary separations, a 2.1 mm Vydac C4 (214TP54) column was used with gradient elution at a flow rate of 100 µl/min. HPLC solvents and system configuration were as described (Tempst et al., 1990), with improved dead volume reduction through the use of glass capillary tubing (Elicone and Tempst, unpublished). Identification of Trpcontaining peptides was done by manual ratio analysis of absorbances at 297 and 277 nm, monitored in real time using an Applied Biosystems model 1000S diode-array detector (Tempst et al., 1990). Fractions were collected by hand, kept on ice for the duration of the run and then stored at -70° C. before repurification and/or analysis. An enzyme blank was done on an equally sized strip of nitrocellulose cut from a blank area of the same blot. Repurifications (second dimension LC) were carried out on a 1.0 mm SGE ODS-2 C18 column using the same solvent system but at a flow rate of 30 µl/min. (C. Elicone, M. Lui, S. Geromanos, H. Erdjument-Bromage, P. Tempst, in press). Samples were always acidified (20% TFA final concentration) and then diluted twofold with 0.1% TFA before rechromatography.

Sequences of 23 peptides separated in this fashion were determined by a combination of automatic Edman degradation, matrix-assisted laser desorption massspectroscopy, and UV spectroscopy.

Peak fractions over background were analyzed by a combination of automated Edman degradation and matrixassisted laser-desorption (MALDI-TOF) mass spectrometry (Geromanos et al., 1994; Elicone et al., 1994). After storage, column fractions were supplemented with neat TFA (to give cleaved by adding an equal volume of thrombin cleavage 55 a final concentration of 10%) before loading onto the sequencer disc and mass spectrometer probe tips. Peptide mass analysis (on 2% aliquots) was carried out using a model LaserTec ResearcH MALDI-TOF instrument (Vestec), with a 337 nm output nitrogen laser and 1.2 m 60 flight tube. The matrix was a-cyano-4-hydroxy cinnamic acid, and a 28 kV ion acceleration and 4.3 kV multiplier voltage were used. Laser power and number of acquisitions were adjusted as judged from optimal deflections of specific maxima, using a Tektronix TDS 520 digitizing oscilloscope. M/z (mass to charge) spectra were generated from the time-of-flight files using GRAMS data analysis software. Every sample was analyzed twice, in the presence and

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absence of a calibrant (25 femtomoles APID), as described (Geromanos et al., 1994). Chemical sequencing (on 95% of the sample) was done using a model 477A instrument from Applied Biosystems (AB). Stepwise liberated PTH-amino acids were identified using an "on-line" 120A HPLC system (AB) equipped with a PTH C18 (2.1×220 mm; 5 micron particle size) column (AB). Instruments and procedures were optimized for femtomole level phenylthiohydantoin amino acid analysis as described (Tempst and Riviere, 1990; Erdjument-Bromage et al., 1993).

Peptide average isotopic masses were summed from the identified residues (including the presumed ones) using ProComp version 1.2 software (obtained from Dr. P. C. Andrews, University of Michigan, Ann Arbor, Mich.). Peptide sequences were compared to entries in various sequence 15 databases using the National Center for Biotechnology Information (NCBI) BLAST program (Altschul et al. 1990). Lower stringency alignments between all peptides and selected proteins were done using the Lipman-Pearson algorithm, available in the 'Lasergene' software package 20 (DNASTAR).

Several protein sequence databases (PIR, SwissProt, translated Genbank) were searched for sequences that match any of the 23 peptide sequences obtained from microsequencing of RAFT1. While sequence similarities with hundreds of different proteins were obtained for many of the 23 peptides, none perfectly matched with any of the entries in the databases, nor did any protein match more than one or two peptides, other than the yeast proteins TOR1 and TOR2 (Kunz et al., 1993). Strikingly, sixteen out of the 23 peptides of RAFT1 could be aligned with the yeast TOR sequences, with varying degrees of similarity (FIG. 4).

#### Example 5

### Molecular Cloning of RAFT1

To generate a probe for isolating a RAFT1 cDNA two degenerate oligonucleotides were used in a mixed oligonucleotide polymerase chain reaction (PCR) (Gould et al, 1989) with rat brain cDNA as template. The sense primer 40 was made to a peptide sequence (TYDPNQP, SEQ ID NO:7) obtained from microsequencing of RAFT1, while the antisense primer corresponds to a sequence (HIDFGD, SEQ ID NO:8) conserved between TOR1, TOR2, and p110 PI-3 Kinase. From the alignments of the RAFT1 peptides to the 45 TORs, this sequence was expected to be 220 amino acids downstream of that encoded by the sense primer. The predicted 660 bp PCR product was obtained, cloned, and its authenticity was verified by DNA sequencing, which showed that it encoded two other sequenced tryptic peptides. 50 The PCR product was, therefore, used as a probe (3' probe) to screen a rat striatum cDNA library, which yielded a 5.5 kb partial cDNA clone. An antisense oligonucleotide to the extreme 5' end of this cDNA was then used in a PCR reaction with a degenerate sense oligonucleotide to another 55 peptide sequence (NDQVFE, SEQ ID NO:9) obtained from microsequencing. The predicted 1.1 kb PCR product was obtained, cloned and used as probe (5' probe) to screen a rat brainstem cDNA library in parallel with the original 3' probe. Phage plaques that hybridized with both probes were 60 isolated and one was found to carry a 8.6 kb insert. A degenerate sense oligonucleotide corresponding to the amino acid sequence TYDPNQP, which was obtained from microsequencing of RAFT1 and aligns to residues 2086 to 2093 of TOR2, and a degenerate antisense primer corre- 65 sponding to amino acids 2296 to 2301 (HIDFGD, SEQ ID NO:8) of TOR2 were used in a PCR reaction with rat whole

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brain cDNA as template. The protocol for the PCR was: an initial 5 min at 94° C., followed by 35 cycles of 94° C. for 40s, 56° C. for 1 min, 72° C. for 1 min, and a final incubation at 72° C. for 5 min. The PCR products were fractionated on a 1.1% agarose gel, the expected 700 bp DNA fragment purified and subcloned into pBluescript.

The RAFT-1 cDNA fragment in pBluescript was amplified by PCR, the product gel purified and labeled by nick translation with a commercial kit (Boehringer Mannheim). This probe (designated 3' probe) was used to screen 1×10° phage plaques of a rat striatum λ ZAP library (Stratagene), as described (Sambrook et al.). Forty seven positive clones were identified and 10 of them were purified by an additional two rounds of screening. None of the inserts contained a complete open reading frame. The 5' end of the largest insert (5.5kb) was used to design a 18 bp antisense oligonucleotide (3.1 as) that was used in another PCR reaction with rat whole brain cDNA as template and a degenerate oligonucleotide corresponding to the amino acid sequence NDQVFE (SEQ ID NO:9 part of a peptide obtained from microsequencing) as the sense primer. The PCR products were fractionated on a 1% agarose gel and a DNA fragment of 1.1 kb isolated and cloned into the vector pCR-II using the TA cloning kit (Invitrogen, San Diego, Calif.). The cDNA fragment was amplified by PCR, the product gel purified and labeled by nick translation. This probe (designated 5' probe) was used to screen 1×10<sup>6</sup> phage plaques from a rat brainstem λ ZAP library. Duplicate filters were screened with the 3' probe. Eight clones hybridized with both the 5' and 3' probes, and five of these were purified through 2 additional rounds of screening. One clone contained a 8.6 kb insert that encodes all 23 peptide sequences obtained by microsequencing.

PCR primer sequences were as follows:

TYDPNQP (SEQ ID NO:7): 5'-GGGGGATCCACNTA (C/T) GA(C/T)CCNAA(C/T) CA(A/G)C-3' (SEQ ID NO:13)

HIDFGD (SEQ ID NO:8): 5'-GCGGAATTC(G/A)
TCNCC(G/A)AA(G/A)TC(T/G/A) AT(G/A)TG-3'
(SEQ ID NO:14)

NDQVFE (SEQ ID NO:9): 5'-GGGGGATCCAA(C/T) GA(C/T)CA(G/A)GTNTT (T/C)GA-3' (SEQ ID NO:15)

3.1as: 5'-GAGCCACCACGATTTGCT-3' (SEQ ID NO:10)

cDNA clones were sequenced using the flourescent terminator method of cycle sequencing on a Applied Biosystems 373a automated DNA sequencer at the DNA analysis Facility of the Johns Hopkins University (Smith et al., 1986; McCombie et al, 1992), or with the dideoxy chain termination method using the Sequenase kit (Amersham, Arlington Heights, Ill.). Oligonucleotides used for sequencing were synthesized on an ABI 394 synthesizer following ABI protocols. DNA sequence data was analyzed using Sequencher software from Gene Codes (Ann Arbor, Mich.). Protein alignments were done with help from the e-mail service of the Computational Biochemistry Research Group (CBRG) at the ETH.

This cDNA contains an open reading frame of 7.6 kb with an initiation methionine codon that conforms to the Kozak consensus sequence (Kozak, 1986) and is preceded by an in-frame termination codon. The protein encoded by this open reading frame contains all 23 peptide sequences obtained by microsequencing of RAFT1 (FIG. 4). Interestingly, none of the peptides sequenced correspond to the C-terminal 250 amino acids of RAFT1, which may indicate that this portion of the protein was proteolytically removed during the purification.

The RAFT1 cDNA predicts a protein of 2550 amino acids with a molecular mass of 289 kDa and a pI of 6.8. Over its entire sequence RAFT1 is 43% identical to TOR2 and 39% identical to TOR1 (FIG. 4). The C-terminal 600 amino acids of RAFT1, which, by analogy to the TORs (Cafferkey et al., 1993; Kunz et al., 1993; Helliwell et al., 1994), is predicted to contain lipid kinase activities, is 65% identical to the yeast proteins. The RAFT1 protein has over 20 consensus sites for phosphorylation by protein kinase C (PKC), including one at serine<sub>2035</sub>, which is in the analogous position to the serine 10 (S<sub>1972</sub> in TOR1 and S<sub>1975</sub> in TOR2) found mutated to arginine in rapamycin resistant yeast (boxed residues in FIG.

The predicted RAFT1 protein is 80 amino acids longer than the TOR proteins, and contains several regions with no 15 apparent homology to the yeast proteins, the largest being a 93 amino acid insertion corresponding to residues 270 to 363 of RAFT1. It is possible that these regions are generated by alternative splicing of exons that may be tissue specific to the brain. They are unlikely to be the translation product 20 Dumont, F. J., Staruch, M. J., Koprak, S. L., Melino, M. R., of unspliced introns because they were found in several cDNA clones isolated from different libraries and the DNA sequence does not reveal consensus splice junction sites.

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#### SEQUENCE LISTING

- (1) GENERAL INFORMATION:
  - (iii) NUMBER OF SEQUENCES: 15
- (2) INFORMATION FOR SEQ ID NO:1:

  - (i) SEQUENCE CHARACTERISTICS:

    (A) LENGTH: 8598 base pairs
    (B) TYPE: nucleic acid
    (C) STRANDEDNESS: double

    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE:

    - (A) ORGANISM: Rattus rattus (G) CELL TYPE: pheochromocytoma (H) CELL LINE: PC12

GGCACGAGCG GCACGAGGCG GTAGCTGAGG CGGTGGCCGA AGCCGCGCGA ACCTCAGGGC

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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GCAGCCAAGG	AGCTCCAGCA	CTATGTCACC	ATGGAACTTC	GAGAGATGAG	TCAGGAGGAG	240
TCTACTCGCT	TCTATGACCA	GCTGAACCAT	CACATTTTTG	AACTGGTTTC	CAGCTCAGAC	300
GCCAATGAGA	GGAAGGGTGG	CATCTTGGCC	ATTGCCAGCC	TCATTGGAGT	GGAAGGTGGG	360
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CGAATCCACG	GGGCCTTGCT	GATCCTCAAC	GAGCTCGTCC	GAATCAGCAG	CATGGAGGGA	900
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TGCAAAGACC	TAATGGGCTT	TGGGACAAAG	CCTCGGCACA	TCACTCCCTT	CACCAGCTTC	1020
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				-conti	nuea	
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CTCCAGGATA CACTCAGAGT CCTCACCTTG TGGTTTGATT ATGGTCACTG GCCAGATGTC	5820
AATGAAGCCC TGGTGGAAGG GGTGAAGGCC ATACAGATTG ACACTTGGTT ACAGGTTATA	5880
CCTCAGCTCA TTGCAAGAAT TGACACGCCC AGACCCTTGG TGGGCCGGCT CATTCACCAG	5940
CTCCTCACAG ATATTGGTCG GTACCACCCA CAGGCCCTCA TCTACCCCCT GACGGTGGCT	6000
TCTAAGTCTA CCACCACAGC CCGTCACAAT GCAGCCAACA AGATCCTGAA GAACATGTGC	6060
GAGCACAGCA ACACGCTAGT ACAGCAGGCC ATGATGGTGA GTGAAGAGCT GATTCGAGTA	6120
GCCATCCTCT GGCATGAGAT GTGGCATGAA GGCCTAGAAG AGGCCTCTCG CTTGTACTTT	6180
GGGGAGAGA ACGTCAAAGG CATGTTTGAG GTGCTGGAGC CCCTGCATGC TATGATGGAA	6240

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CGCGGTCCCC	AGACCCTGAA	GGAAACGTCC	TTTAATCAGG	CATATGGTCG	AGATTTAATG	6300
GAGGCACAAG	AATGGTGCCG	AAAGTACATG	AAATCAGGGA	ACGTCAAGGA	CCTCACCCAA	6360
GCCTGGGACC	TCTACTATCA	CGTGTTCAGA	CGGATCTCCA	AGCAGCTACC	ACAGCTCACA	6420
TCCCTGGAGC	TGCAGTATGT	GTCCCCCAAA	CTTTTGATGT	GCAGAGACCT	TGAATTGGCT	6480
GTGCCAGGAA	CATATGACCC	CAACCAGACA	ATCATTCGCA	TTCAGTCCAT	AGCCCCGTCT	6540
TTGCAAGTCA	TCACATCCAA	GCAGAGGCCT	CGGAAGCTGA	CCCTGATGGG	CAGCAATGGG	6600
CACGAGTTTG	TTTTCCTCCT	GAAAGGCCAT	GAAGATCTGC	GGCAGGACGA	GCGAGTGATG	6660
CAGCTCTTTG	GCCTGGTGAA	CACACTCCTA	GCCAATGACC	CAACTTCTCT	TCGAAAGAAC	6720
CTCAGCATCC	AGAGATACGC	CGTCATTCCT	CTGTCCACCA	ACTCGGGCCT	GATTGGCTGG	6780
GTGCCCCACT	GTGACACGCT	GCATGCCCTC	ATCCGGGACT	ACAGAGAGAA	GAAGAAGATC	6840
CTGCTGAACA	TCGAGCACCG	CATCATGCTG	CGGATGGCTC	CTGACTATGA	CCACCTGACT	6900
CTGATGCAGA	AGGTGGAGGT	GTTTGAGCAT	GCTGTCAACA	ACACAGCCGG	GGATGACCTG	6960
GCCAAGCTGC	TGTGGCTGAA	AAGCCCCAGC	TCAGAGGTGT	GGTTTGACCG	AAGAACCAAT	7020
TATACTCGCT	CCCTGGCTGT	CATGTCCATG	GTTGGATACA	TTTTAGGCCT	TGGAGACAGG	7080
CACCCATCCA	ACCTGATGCT	GGACCGGCTG	AGTGGAAAGA	TCCTGCACAT	TGACTTTGGG	7140
GACTGCTTTG	AGGTTGCTAT	GACCAGAGAG	AAATTTCCAG	AAAAGATTCC	ATTTAGACTA	7200
ACAAGAATGT	TGACCAATGC	TATGGAGGTT	ACCGGTCTCG	ATCGCAACTA	TAGAACCACA	7260
TGCCACACAG	TGATGGAGGT	GCTTCGGGAG	CACAAGGACA	GTGTCATGGC	TGTGCTAGAA	7320
GCCTTTGTCT	ATGACCCTCT	GCTGAATTGG	AGGCTGATGG	ACACAAATGC	CAAAGGCAAC	7380
AAGCGGTCCC	GAACCAGGAC	AGACTCCTAT	TCTGCAGGCC	AGTCAGTAGA	AATTTTGGAC	7440
GGTGTAGAAC	TTGGAGAACC	AGCCCATAAG	AAAACAGGGA	CCACTGTGCC	AGAATCCATC	7500
CATTCTTTCA	TTGGAGATGG	TTTGGTGAAA	CCAGAAGCCT	TAAACAAGAA	AGCTATTCAG	7560
ATTATTAACA	GGGTTCGAGA	TAAGCTCACT	GGTCGGGATT	TCTCTCATGA	TGACACTTTG	7620
GATGTTCCAA	CCCAAGTGGA	ACTGCTTATC	AAGCAAGCGA	CATCTCATGA	GAACCTCTGC	7680
CAGTGCTACA	TTGGCTGGTG	TCCTTTCTGG	TAACCAAGGC	CTGGCAAAGA	AAATCATCTC	7740
CTCCGATGCT	TTTGTACCTT	GGTCTGTGCT	TCCAGTGGAC	TGAAACCATG	GTCATAAAGT	7800
TGGACTTTGT	TAATATTTTG	AAATGTATAT	GAAAAGAACT	ACTGTATATT	CAAAGTTGGC	7860
TTATGCCAAC	CTCCTAGCTG	CTGTTGAAAA	GACACTGTCA	GAAACACAAG	GCTTGATTCA	7920
GTTCCCAGGA	CAGTGAAACA	CAGTAATCCT	ACAGAAACCA	AGCCTTTGAT	TTTGGGAGAA	7980
CAGAAGATGA	GTAACTGACT	AAGAAATACG	GGTTTGGACT	TAACTTACAG	AAGAACTCAT	8040
CATACGCATT	TGCTGACCGA	ATAATCTAGT	TGATCCTCTC	AACCAGGGGC	TTCAACAGCA	8100
AGGACACAGA	TGTCAGCACT	CCACCATCCT	GTTACCTCAC	CCGTCCCTGG	ATGCAGTGGC	8160
AACATCTGCA	GGATGGGCCA	CCGTGTGTGT	AAGAAGATCT	GTCTTCCACC	TGATCCCATG	8220
ATGCTGAACC	TCACAGAGCC	GGCCTTCCAG	GAAGGACGTT	TGCTCAGACG	CCTGGCCACC	8280
GAGGATGAGC	AGGTGTGCCA	GGATCTCAGT	GCAGGGTCCA	CGCTGGCCCT	GCTGCTGTGT	8340
TCAGTGAGGG	ATGGATATGT	TGTGTTTGCA	GCAGGGACTC	AGAACACAAA	TGCTTTTGTG	8400
GAAGTGCTGA	TCTCAGAGGG	ACACTAGCGC	AGGTTGTGAA	TTAAGAGCAA	AGTAAATATC	8460
CAACTAAACA	CAAAGTATAA	GTGAAGCCAC	ATCTAGACAC	CATTGTATCT	GAGTAATTTT	8520
TGTGCCAATA	AATGACATCA	GAATTTTAAA	AGTAAAAAAA	ACGATATCAA	GCTTATCGAT	8580
ACCGTCGACC	TCGAGGGG					8598

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(2) INFORMATION FOR SEQ ID NO:2:

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ν		i) S	EQUEI	NCE ( LENG' LYPE	CHARI TH: :	ACTE		ics:	cids						
	(D) TOPOLOGY: linear  (ii) MOLECULE TYPE: protein														
	(ii) MOLECULE TYPE: protein														
	(iii) HYPOTHETICAL: YES														
(iv) ANTI-SENSE: NO															
<ul><li>(vi) ORIGINAL SOURCE:</li><li>(A) ORGANISM: Rattus rattus</li><li>(F) TISSUE TYPE: pheochromocytoma</li><li>(G) CELL TYPE: PC12</li></ul>															
(G) CELL TYPE: PC12  (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:															
Met Leu Gly Thr Gly Pro Ala Thr Ala Thr Ala Gly Ala Ala Thr Ser  1 5 10 15  Ser Asn Val Ser Val Leu Gln Gln Phe Ala Ser Gly Leu Lys Ser Arg															
Ser	Asr	val	. Ser 20	· Val	. Leu	Glr	Gln	Phe 25	Ala	Ser	Gly	Leu	Lys 30	Ser	Arg
Asn	Glu	35	Thr	Arg	Ala	Lys	Ala 40	Ala	Lys	Glu	Leu	Gln 45	His	Tyr	Val
Thr	Met 50	Glu	Leu	Arg	Glu	Met 55	Ser	Gln	Glu	Glu	Ser 60	Thr	Arg	Phe	Tyr
65					70					75					Ala 80
				85					90					95	Val
			100					105					110		Arg
		115					Pro 120					125			
	130					135	Ala				140				
145					150		Ala			155					160
				165			Ala		170					175	
			180				Phe	185					190		
		195					Asp 200					205			
Ala	Val 210	Ala	Ala	Leu	Arg	Ala 215	Сув	Leu	Ile	Leu	Thr 220	Thr	Gln	Arg	Glu
225					230		Gln			235					240
Ala	Glu	Lys	Gly	Phe 245	Asp	Glu	Thr	Leu	Ala 250	Lys	Glu	Lys	Gly	Met 255	Asn
			260				Ala	265					270		
Arg		275					280					285			
Ile	Thr 290	Gln	Gln	Gln	Leu	Val 295	His	qaA	Lys	Tyr	аұЭ 008	ayı	Asp	Leu	Met

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# Gly Phe Gly Thr Lys Pro Arg His Ile Thr Pro Phe Thr Ser Phe Gln 305 310 315 320Ala Val Gln Pro Gln Gln Ser Asn Ala Leu Val Gly Leu Leu Gly Tyr 325 330 335 Lys Ser Thr Leu Val Glu Ser Arg Cys Cys Arg Asp Leu Met Glu Glu 355 360 365 Asn Ser Leu Ile Gln Met Thr Ile Leu Asn Leu Leu Pro Arg Leu Val 385 390 395 400 Ala Phe Arg Pro Ser Ala Phe Thr Asp Thr Gln Tyr Leu Gln Asp Thr 405 410 415Met Asn His Val Leu Ser Cys Val Lys Lys Glu Lys Glu Arg Thr Ala 420 425 430 Ala Phe Gln Ala Leu Gly Leu Leu Ser Val Ala Val Arg Ser Glu Phe $435 \ \ \, 440 \ \ \, 445$ Lys Val Tyr Leu Pro Arg Val Leu Asp Ile Ile Arg Ala Ala Leu Pro $450 \hspace{1.5cm} 455 \hspace{1.5cm} 460 \hspace{1.5cm}$ Pro Lys Asp Phe Ala His Lys Arg Gln Lys Thr Val Gln Val Asp Ala 465 $\phantom{\bigg|}470\phantom{\bigg|}470\phantom{\bigg|}475\phantom{\bigg|}$ Thr Val Phe Thr Cys Ile Ser Met Leu Ala Arg Ala Met Gly Pro Gly 485 490 495Ile Gln Gln Asp Ile Lys Glu Leu Leu Glu Pro Met Leu Ala Val Gly 500 500 510 Leu Ser Pro Ala Leu Thr Ala Val Leu Tyr Asp Leu Ser Arg Gln Ile 515 520 525Pro Gln Leu Lys Lys Asp Ile Gln Asp Gly Leu Leu Lys Met Leu Ser 530 540Leu Val Leu Met His Lys Pro Leu Arg His Pro Gly Met Pro Lys Gly 545 $^{\circ}$ . 550 $^{\circ}$ 555 $^{\circ}$ 560 Leu Ala His Gln Leu Ala Ser Pro Gly Leu Thr Thr Leu Pro Glu Ala 565 570 575Ser Asp Val Ala Ser Ile Thr Leu Ala Leu Arg Thr Leu Gly Ser Phe $580 \hspace{1.5cm} 585 \hspace{1.5cm} 590$ Glu Phe Glu Gly His Ser Leu Thr Gln Phe Val Arg His Cys Ala Asp $595 \hspace{1.5cm} 600 \hspace{1.5cm} 605 \hspace{1.5cm}$ His Phe Leu Asn Ser Glu His Lys Glu Ile Arg Met Glu Ala Ala Arg 610 $\,$ 615 $\,$ 620 $\,$ Thr Cys Ser Arg Leu Leu Thr Pro Ser Ile His Leu Ile Ser Gly His 625 $\phantom{\bigg|}$ 630 $\phantom{\bigg|}$ 635 $\phantom{\bigg|}$ 640 Ala His Val Val Ser Gln Thr Ala Val Gln Val Val Ala Asp Val Leu 645 650 655Ser Lys Leu Leu Val Val Gly Ile Thr Asp Pro Asp Pro Asp Ile Arg 660 665 670 Tyr Cys Val Leu Ala Ser Leu Asp Glu Arg Phe Asp Ala His Leu Ala 675 680 685Phe Glu Ile Arg Glu Leu Ala Ile Cys Thr Val Gly Arg Leu Ser Ser 705 710 715 720

Met Asn Pro Ala Phe Val Met Pro Phe Leu Arg Lys Met Leu Ile Gln

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				725	,				730					735	
Ile	e Leu	Thr	740	Lev	ı Glu	His	Ser	Gly 745	Ile	Gly	Arg	Ile	Lys 750		Gln
Ser	Ala	755	Met	: Leu	Gly	His	Leu 760	Val	Ser	Asn	Ala	Pro 765		Leu	Ile
Arg	770	Tyr	Met	: Glu	Pro	Ile 775	Leu	Lys	Ala	Leu	Ile 780	Leu	Lys	Leu	Lys
Asp 785	Pro	Asp	Pro	Asp	790		Pro	Gly	Val	Ile 795	Asn	Asn	Val	Leu	Ala 800
Thr	Ile	Gly	Glu	Leu 805	Ala	Gln	Val	Ser	Gly 810		Glu	Met	Arg	Lys 815	Trp
Val	Asp	Glu	Leu 820		Val	Ile	Ile	Met 825	Asp	Met	Leu	Gln	Asp 830	Ser	Ser
Leu	Leu	Ala 835	Lys	Arg	Gln	Val	Ala 840	Leu	Trp	Thr	Leu	Gly 845	Gln	Leu	Val
Ala	Ser 850	Thr	Gly	Tyr	Val	Val 855	Glu	Pro	Tyr	Arg	<b>Lys</b> 860	Tyr	Pro	Thr	Leu
Leu 865	Glu	Val	Leu	Leu	Asn 870	Phe	Leu	Lys	Thr	Glu 875	Gln	Asn	Gln	Gly	Thr 880
Arg	Arg	Glu	Ala	Ile 885	Arg	Val	Leu	Gly	Leu 890	Leu	Gly	Ala	Leu	Asp 895	Pro
Tyr	Lys	His	Lys 900	Val	Asn	Ile	Gly	Met 905	Ile	Asp	Gln	Ser	Arg 910	Asp	Ala
Ser	Ala	Val 915	Ser	Leu	Ser	Glu	Ser 920	Lys	Ser	Ser	Gln	Asp 925	Ser	Ser	Asp
Tyr	Ser 930	Thr	Ser	Glu	Met	Leu 935	Val	Asn	Met	Gly	Asn 940	Leu	Pro	Leu	Asp
Glu 945	Phe	Tyr	Pro	Ala	Val 950	Ser	Met	Val	Ala	Leu 955	Met	Arg	Ile	Phe	Arg 960
Asp	Gln	Ser	Leu	Ser 965	His	His	His	Thr	Met 970	Val	Val	Gln	Ala	Ile 975	Thr
Phe	Ile	Phe	<b>Lys</b> 980	Ser	Leu	Gly	Leu	<b>Lys</b> 985	Сув	Val	Gln	Phe	Leu 990	Pro	Gln
Val	Met	Pro 995	Thr	Phe	Leu	Asn	Val 1000		Arg	Val	Сув	Asp 1005		Ala	Ile
Arg	Glu 1010	Phe )	Leu	Phe	Gln	Gln 1015	Leu	Gly	Met	Leu	Val 1020		Phe	Val	Lys
Ser 1025	His 5	Ile	Arg	Pro	Tyr 1030		Asp	Glu	Ile	Val 1035		Leu	Met	Arg	Glu 1040
Phe	Trp	Val		Asn 1045	Thr	Ser	Ile		Ser 1050		Ile	Ile	Leu	Leu 1055	Ile
Glu	Gln	Ile	Val 1060		Ala	Leu	Gly	Gly 1065		Phe	Lys	Leu	Tyr 1070		Pro
Gln	Leu	Ile 1075	Pro	His	Met	Leu	Arg 1080		Phe	Met	His	Asp 1085		Ser	Gln
Gly	Arg 1090		Val	Ser	Ile	Lys 1095		Leu	Ala	Ala	lle 1100		Leu	Phe	Gly
Ala 1105		Leu	qaA	qaA	Tyr 1110		His	Leu	Leu	Leu 1115		Pro	Ile	Val	Lys 1120
Leu	Phe	Asp	Ala	Pro 1125	Glu	Val	Pro	Leu	Pro 1130		Arg	Lys	Ala	Ala 1135	
Glu	Thr	Val	Asp 1140		Leu	Thr		Ser 1145		Asp	Phe		Asp 1150		Ala

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Ser	Arg	Ile 115	Ile 5	His	Pro	Ile	Val 116	Arg 0	Thr	Leu	Asp	Gln 116	Ser 5	Pro	Glu
Leu	Arg 117	Ser 0	Thr	Ala	Met	Asp 117	Thr 5	Leu	Ser	Ser	Leu 118		Phe	Gln	Leu
Gly 118	- <b>Ly</b> в 5	Lys	Tyr	Gln	Ile 119	Phe	Ile	Pro	Met	Val 119		Lys	Val	Leu	Val 1200
Arg	His	Arg	Ile	120	His 5	Gln	Arg	Tyr	Asp 121		Leu	Ile	Cys	Arg 121	
Val	Lys	Gly	Tyr 122	Thr 0	Leu	Ala	Asp	Glu 122	Glu 5	Glu	Asp	Pro	Leu 123		Tyr
Gln	His	Arg 123	Met 5	Leu	Arg	Ser	Ser 124		Gly	Asp	Ala	Leu 124	Ala 5	Ser	Gly
Pro	Val 125	Glu 0	Thr	Gly	Pro	Met 125		Lys	Leu	His	Val 126		Thr	Ile	Asn
Leu 126	Gln 5	Lys	Ala	Trp	Gly 1270	Ala )	Ala	Arg	Arg	Val 127		Lys	Asp	Asp	Trp 1280
Leu	Glu	Trp	Leu	Arg 1285	Arg	Leu	Ser	Leu	Glu 129		Leu	Lys	Asp	Ser 129	
Ser	Pro	Ser	Leu 130	Arg 0	Ser	Сув	Trp	Ala 1305	Leu	Ala	Gln	Ala	Tyr 131		Pro
Met	Ala	Arg 1315	Asp	Leu	Phe	Asn	Ala 1320	Ala	Phe	Val	Ser	Cys 1325		Ser	Glu
Leu	Asn 1330	Glu )	Asp	Gln	Gln	Asp 1335	Glu 5	Leu	Ile	Arg	Ser 1340	Ile )	Glu	Leu	Ala
Leu 1345	Thr	Ser	Gln	Asp	Ile 1350	Ala	Glu	Val	Thr	Gln 1355		Leu	Leu	Asn	Leu 1360
Ala	Glu	Phe	Met	Glu 1365	His	Ser	Asp	Lys	Gly 1370	Pro	Leu	Pro	Leu	Arg 1375	
			1380					1385	i				1390	)	
		1395		Leu			1400					1405	<b>i</b>		
Thr	Pro 1410	Ala	Ile	Leu	Glu	Ser 1415	Leu	Ile	Ser	Ile	Asn 1420		Lys	Leu	Gln
1425	;				1430					1435					1440
				Ile 1445					1450					1455	
Glu	Asp	Ala	Leu 1460	Val	Ala	Tyr	Asp	Lys 1465	Lys	Met	Asp	Thr	Asn 1470		Asp
Asp	Pro	Glu 1475	Leu	Met	Leu	Gly	Arg 1480	Met	Arg	Сув		Glu 1485		Leu	Gly
Glu	Trp 1490	Gly	Gln	Leu		Gln 1495		Cys	Cys		Lys 1500		Thr	Leu	Val
Asn 1505	Asp	Glu	Thr	Gln .	Ala : 1510	Lys	Met .	Ala.		Met 1515	Ala	Ala	Ala	Ala	Ala 1520
Trp	Gly	Leu		Gln ' 1525	Trp .	qaA	Ser		Glu 1530		Tyr	Thr		Met 1535	
Pro	Arg .		Thr 1540	His !	Asp (	Gly .		Phe ' 1545	Tyr	Arg .	Ala		Leu 1550		Leu
His	Gln .	Asp 1555	Leu	Phe :	Ser 1		Ala ( 1560	Gln (	Gln	Сув		Авр 1565	Lys	Ala	Arg

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Asp	Leu	Leu	Asp	Ala	Glu	Leu	Thr	Ala	Met	Ala	Glv	Glu	Ser	Tur	Ser
	157					157					158			-71	561
Arg 158	Ala 5	Tyr	Gly	Ala	Met 159		Ser	Cys	His	Met 159		Ser	Glu	Leu	Glu 1600
Glu	Val	Ile	Gln	Tyr 160	Lys 5	Leu	Val	Pro	Glu 161		Arg	Glu	Ile	Ile 161	
Gln	Ile	Trp	Trp 162	Glu 0	Arg	Leu	Gln	Gly 162	Cys 5	Gln	Arg	Ile	Val 163		Asp
Trp	Gln	Lys 163	Ile 5	Leu	Met	Val	Arg 164		Leu	Val	Val	Ser 164		His	Glu
Asp	Met 1650	Arg )	Thr	Trp	Leu	Lys 165	<b>Ty</b> r 5	Ala	Ser	Leu	Cys 166		Lys	Ser	Gly
Arg 166	Leu 5	Ala	Leu	Ala	His 167		Thr	Leu	Val	Leu 167		Leu	Gly	Val	Asp 1680
Pro	Ser	Arg	Gln	Leu 1689		His	Pro	Leu	Pro 169		Val	His	Pro	Gln 169	
Thr	Tyr	Ala	Tyr 170	Met 0	Lув	Asn	Met	Trp		Ser	Ala	Arg	Lys 171		Asp
Ala	Phe	Gln 171	His	Met	Gln	His	Phe 172	Val	Gln	Thr	Met	Gln 172		Gln	Ala
Gln	His 1730		Ile	Ala	Thr	Glu 173		Gln	Gln	His	Lys 174		Glu	Leu	His
Lys 1745	Leu	Met	Ala	Arg	C <b>y</b> s 1750	Phe	Leu	Lys	Leu	Gly 175		Trp	Gln	Leu	Asn 1760
Leu	Gln	Gly	Ile	Asn 1765	Glu	Ser	Thr	Ile	Pro		Val	Leu	Gln	Tyr 1775	
Ser	Ala	Ala	Thr 178	Glu )	His	qaA	Arg	Ser 1785	Trp	Tyr	Lys	Ala	Trp		Ala
Trp	Ala	Val 1795		Asn	Phe	Glu	Ala 180		Leu	His	Tyr	<b>Lys</b> 1805		Gln	Asn
Gln	Ala 1810	Arg	Asp	Glu	Lys	Lys 1815	Lys	Leu	Arg	His	Ala 1820		Gly	Ala	Asn
Ile 1825	Thr	Asn	Ala	Thr	Thr 1830		Ala	Thr	Thr	Ala 1835		Ser	Ala	Ala	Ala 1840
Ala	Thr	Ser	Thr	Glu 1845		Ser	Asn	Ser	Glu 1850		Glu	Ala	Glu	Ser 1855	
Glu	Ser	Ser	Pro 1860	Thr	Pro	Ser	Pro	Leu 1865		Lys	Lys	Val	Thr 1870		Asp
Leu	Ser	Lys 1875	Thr	Leu	Leu	Leu	<b>Tyr</b> 1880		Val	Pro	Ala	Val 1885		Gly	Phe
Phe	Arg 1890	Ser	Ile	Ser	Leu	Ser 1895	Arg	Gly	Asn	Asn	Leu 1900		Asp	Thr	Leu
Arg 1905	Val	Leu	Thr	Leu	Trp 1910	Phe	Asp	Tyr	Gly	His 1915		Pro	Asp	Val	Asn 1920
Glu	Ala	Leu	Val	Glu 1925		Val	Lys	Ala	Ile 1930		Ile	Asp	Thr	Trp 1935	
Gln	Val	Ile	Pro 1940	Gln	Leu	Ile	Ala	Arg 1945		Asp	Thr	Pro	Arg 1950		Leu
Val	Gly :	Arg 1955	Leu	Ile	His	Gln	Leu 1960	Leu	Thr	Asp		Gly 1965		Tyr	His
	Gln 1 1970	Ala	Leu	Ile		Pro 1975		Thr	Val		Ser 1980		Ser	Thr	Thr
Thr .	Ala	Arg	His	Asn .	Ala	Ala	Asn	Lys	Ile	Leu	Lys	Asn	Met	Cys	Glu

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2000

1995

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1990

1985

198	5				199	U				199	5				2000
His	Ser	Asn	Thr	Leu 200		Gln	Gln	Ala	Met 201		Val	Ser	Glu	Glu 201	Leu 5
Ile	Arg	Val	Ala 202	Ile O	Leu	Trp	His	Glu 202	Met 5	Trp	His	Glu	Gly 203		Glu
Glu	Ala	Ser 203	Arg 5	Leu	Tyr	Phe	Gly 2040		Arg	Asn	Val	Lys 204		Met	Phe
Glu	Val 205	Leu 0	Glu	Pro	Leu	His 2055	Ala	Met	Met	Glu	Arg 206		Pro	Gln	Thr
Leu 206	Lys 5	Glu	Thr	Ser	Phe 207	Asn 0	Gln	Ala	Tyr	Gly 207	Arg 5	Asp	Leu	Met	Glu 2080
Ala	Gln	Glu	Trp	Cys 208!		Lys	Tyr	Met	Lys 209		Gly	Asn	Val	<b>Lys</b> 209	
Leu	Thr	Gln	Ala 210		Asp	Leu	Tyr	Tyr 2105		Val	Phe	Arg	Arg 211		Ser
Lys	Gln	Leu 211	Pro	Gln	Leu	Thr	Ser 2120		Glu	Leu	Gln	Tyr 2125		Ser	Pro
Lys	Leu 213		Met	Cys	Arg	Asp 2135		Glu	Leu	Ala	Val 2140		Gly	Thr	Tyr
Asp 214	Pro 5	Asn	Gln	Thr	Ile 215	Ile )	Arg	Ile	Gln	Ser 215		Ala	Pro	Ser	Leu 2160
Gln	Val	Ile	Thr	Ser 2165	Lys 5	Gln	Arg	Pro	Arg 2170	Lys )	Leu	Thr	Leu	Met 2175	
Ser	Asn	Gly	His 2180		Phe	Val	Phe	Leu 2185		Lys	Gly	His	Glu 2190		Leu
Arg	Gln	Asp 2195	Glu	Arg	Val	Met	Gln 2200		Phe	Gly	Leu	Val 2205		Thr	Leu
Leu	Ala 2210	Asn )	Asp	Pro	Thr	Ser 2215	Leu	Arg	Lys	Asn	Leu 2220		Ile	Gln	Arg
Tyr 222!	Ala	Val	Ile	Pro	Leu 2230	Ser	Thr	Asn	Ser	Gly 2235		Ile	Gly	Trp	Val 2240
Pro	His	Cys	Asp	Thr 2245	Leu	His	Ala	Leu	Ile 2250		Asp	Tyr	Arg	Glu 2255	
Lys	Lys	Ile	Leu 2260		Asn	Ile		His 2265		Ile	Met	Leu	Arg 2270		Ala
Pro	Asp	Tyr 2275		His	Leu	Thr	Leu 2280		Gln	Lys	Val	Glu 2285		Phe	Glu
His	Ala 2290	Val	Asn	Asn	Thr	Ala 2295	Gly	qaA	Asp		Ala 2300		Leu	Leu	Trp
Leu 2305	Lys	Ser	Pro	Ser	Ser 2310	Glu	Val	Trp	Phe	Asp 2315	Arg	Arg	Thr	Asn	Tyr 2320
Thr	Arg	Ser	Leu	Ala 2325		Met	Ser		Val 2330		Tyr	Ile	Leu	Gly 2335	
Gly	Asp	Arg	His 2340	Pro	Ser	Asn		Met 2345	Leu	Asp	Arg		Ser 2350		Lys
Ile	Leu	His 2355		Asp	Phe	Gly	Asp 2360		Phe	Glu		Ala 2365	Met	Thr	Arg
Glu	<b>Lу</b> в 2370		Pro	Glu		Ile 2375		Phe	Arg		Thr 2380		Met	Leu	Thr
Asn 2385		Met .	Glu		Thr 2390	Gly	Leu .	Asp		Asn 2395		Arg	Thr		Cys 2400
His	Thr	Val	Met	Glu 2405	Val	Leu .	Arg		His 2410		Asp	Ser		Met 2415	Ala

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Val Leu Glu Ala Phe Val Tyr Asp Pro Leu Leu Asn Trp Arg Leu Met 2420 2425 2430

Asp Thr Asn Ala Lys Gly Asn Lys Arg Ser Arg Thr Arg Thr Asp Ser 2435 2445

Glu Pro Ala His Lys Lys Thr Gly Thr Thr Val Pro Glu Ser Ile His 2465 2470 2475 2480

Ser Phe Ile Gly Asp Gly Leu Val Lys Pro Glu Ala Leu Asn Lys Lys 2495 2495

Ala Ile Gln Ile Ile Asn Arg Val Arg Asp Lys Leu Thr Gly Arg Asp 2500 2505 Leu Thr Gly Arg Asp

Phe Ser His Asp Asp Thr Leu Asp Val Pro Thr Gln Val Glu Leu Leu 2515 2520 2525

Ile Lys Gln Ala Thr Ser His Glu Asn Leu Cys Gln Cys Tyr Ile Gly 2530 2540

Trp Cys Pro Phe Trp

- (2) INFORMATION FOR SEQ ID NO:3:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 2470 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Saccharomyces cerevisiae
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Met Glu Pro His Glu Glu Gln Ile Trp Lys Ser Lys Leu Leu Lys Ala 1  $\phantom{\bigg|}$  5  $\phantom{\bigg|}$  10  $\phantom{\bigg|}$  15

Ala Asn Asn Asp Met Asp Met Asp Arg Asn Val Pro Leu Ala Pro Asn 20 25 30

Leu Asn Val Asn Met Asn Met Lys Met Asn Ala Ser Arg Asn Gly Asp 35 40 45

Glu Phe Gly Leu Thr Ser Ser Arg Phe Gly Gly Val Val Ile Gly Ser 50

Leu Thr Ser Asp Tyr Lys Glu Glu Arg Lys Leu Ala Ser Ile Ser Leu 85 90 95

Phe Asp Leu Leu Val Ser Leu Glu His Glu Leu Ser Ile Glu Glu Phe 100 \$100\$

Gln Ala Ile Ser Asn Asp Ile Asn Asn Lys Ile Leu Glu Leu Val His 115 \$120\$

Leu Ile Ser Phe Tyr Ala Tyr Thr Glu Arg Leu Pro Asn Glu Thr Ser 145 150 155 160

Arg Leu Ala Gly Tyr Leu Arg Gly Leu Ile Pro Ser Asn Asp Val Glu 165 170 175

Val Met Arg Leu Ala Ala Lys Thr Leu Gly Lys Leu Ala Val Pro Gly 180 185 190

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											_	-001	1011	ruec	
Gly	7 Th:	r Tyr 195	Thi	Ser	Asp	Phe	e Val		ı Phe	e Glu	Ile	Lys 205		r Cy	s Leu
Glu	210	p Lev	Thr	Ala	a Ser	Th:		Ly:	s Asr	n Ser	Phe 220		Sei	r Sei	r L <b>y</b> s
Pro 225	As <sub>l</sub>	His	Ala	Lys	His 230		a Ala	Let	ı Lev	1le 235		Thr	Ala	a Let	1 Ala 240
Glu	Ası	n Cys	Pro	Tyr 245		Lev	ı Tyr	Glr	Tyr 250		Asn	Ser	Tle	255	ı Asp
Asn	Ile	e Trp	Arg 260	Ala	Leu	Arg	J Asp	Pro 265		Leu	Val	Ile	Arg 270		qaA e
Ala	Ser	11e 275	Thr	Leu	Ala	Lys	280	Leu	Ser	Thr	Leu	Arg 285		Arg	Asp
Pro	Glr 290	Leu	Thr	Ser	Gln	Trp 295	Val	Gln	Arg	Leu	Ala 300	Thr	Ser	Сув	Glu
Tyr 305	Gly	Phe	Gln	Val	Asn 310	Thr	Leu	Glu	Cys	Ile 315	His	Ala	Ser	Leu	Leu 320
Val	Tyr	Lys	Glu	Ile 325		Phe	Leu	Lys	930 088	Pro	Phe	Leu	Asn	Gln 335	
Phe	Asp	Gln	Met 340	Cys	Leu	Asn	Cys	Ile 345		Tyr	Glu	Asn	His 350		Ala
Lys	Met	11e 355	Arg	Glu	Lys	Ile	Tyr 360	Gln	Ile	Val	Pro	Leu 365	Leu	Ala	Ser
Phe	Asn 370	Pro	Gln	Leu	Phe	Ala 375		Lys	Tyr	Leu	His 380	Gln	Ile	Met	Asp
Asn 385	Tyr	Leu	Glu	Ile	Leu 390	Thr	Asn	Ala	Pro	Ala 395	Lys	Lys	Ile	Pro	His 400
Leu	Lys	Авр	Asp	Lys 405	Pro	Gln	Ile	Leu	Ile 410	Ser	Ile	Gly	qaA	Ile 415	Ala
Tyr	Glu	Val	Gly 420	Pro	Asp	Ile	Ala	Pro 425	Tyr	Val	Lys	Gln	Ile 430	Leu	Asp
		435					440			Lys		445			
Glu	Asn 450	Glu	Ile	Phe	Tyr	Cys 455	Ile	Gly	Arg	Leu	Ala 460	Val	Pro	Leu	Gly
Pro 465	Val	Leu	Gly	Lys	Leu 470	Leu	Asn	Arg	Asn	Ile 475	Leu	Asp	Leu	Met	Phe 480
Lys	Cys	Pro	Leu	Ser 485	Asp	Tyr	Met	Gln	Glu 490	Thr	Phe	Gln	Ile	Leu 495	Thr
			500					505		Asn			510		
Leu	Val	Cys 515	Ser	Thr	Leu	Ser	Gly 520	Thr	Pro	Phe	Ile	Gln 525	Pro	Gly	Ser
Pro	Met 530	Glu	Ile	Pro		Phe 535	Ser	Arg	Glu	Arg	Ala 540	Arg	Glu	Trp	Arg
Asn 545	Lys	Ser.	Ile	Leu	Gln 550	Lys	Thr	Gly	Glu	Ser 555	Asn	Asp	Asp		Asn 560
Asp	Ile	Lys	Ile	Ile 565	Ile	Gln	Ala	Phe	Arg 570	Met	Leu	Lys	Asn	Ile 575	Lys
			580					585		Ile			590		-
		595					600			Leu		605			
Cys	Glu	Ile	Tyr	Val	Lys .	Asp	Asn	Ile	Cys	Lys	Gln	Thr	Ser	Leu	His

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	61	0				615	5				620	)			
Se. 62	r Le	u As	n Th	r Val	L Ser 630	Glu	ı Val	l Leu	se:	r Ly:	s Let	ı Le	u Al	a Il	e Thr 640
11	e Al	a As	p Pro	645	ı Gln	Asp	Ile	e Arç	650		ı Val	l Lei	ı Ly	65	n Leu 5
Ası	n Pro	о Су	s Phe 660	e Asp	Pro	Glr	Leu	1 Ala 665		n Pro	Asp	Ası	67		g Leu
Lei	ı Phe	● Th:	r Ala	ı Lev	His	ĄaĄ	Glu 680	ser	Phe	e Asr	ıle	685		r Val	l Ala
Met	690	ı Let	ı Val	l Gly	Arg	Leu 695	Ser	Ser	Val	l Asr	700		ту	c Val	l Ile
Pro 705	Sei	: Ile	e Arg	l Lys	710	Leu	Leu	Glu	Lev	1 Let 715		Lys	Let	ı Lys	720
Ser	Thr	Ser	: Ser	725	Glu	Lys	Glu	Glu	Thr 730	Ala	Ser	Lev	Let	1 Cys 735	Thr
Lev	ı Ile	Arg	740	Ser	Lys	Asp	Val	Ala 745	Lys	Pro	Tyr	Ile	Glu 750		Leu
Leu	Asn	Val 755	. Leu	Leu	Pro	Lys	Phe 760		Asp	Thr	Ser	Ser 765		· Val	Ala
Ser	770	Ala	Leu	Arg	Thr	Ile 775	Gly	Glu	Leu	Ser	Val 780	Val	Gly	Gly	Glu
Asp 785	Met	Lys	Ile	Tyr	Leu 790	Lys	Asp	Leu	Phe	Pro 795	Leu	Ile	Ile	Lys	Thr 800
Phe	Gln	Asp	Gln	Ser 805	Asn	Ser	Phe	Lys	Arg 810		Ala	Ala	Leu	Lys 815	
Leu	Gly	Gln	Leu 820	Ala	Ala	Ser	Ser	Gly 825	Tyr	Val	Ile	Asp	Pro 830		Leu
Asp	Tyr	Pro 835	Glu	Leu	Leu	Gly	Ile 840	Leu	Val	Asn	Ile	Leu 845	Lys	Thr	Glu
Asn	Ser 850	Gln	Asn	Ile	Arg	Arg 855	Gln	Thr	Val	Thr	Leu 860	Ile	Gly	Ile	Leu
Gly 865	Ala	Ile	Asp	Pro	Tyr 870	Arg	Gln	Lys	Glu	Arg 875	Glu	Val	Thr	Ser	Thr 880
				885	Glu				890					895	
Leu	Met	Gln	Gly 900	Met	Ser	Pro	Ser	Asn 905	Asp	Glu	Tyr	Tyr	Thr 910	Thr	Val
		915			Leu		920					925			
Tyr	His 930	Thr	Ala	Val	Ile	Gln 935	Ala	Ile	Met	His	Ile 940	Phe	Gln	Thr	Leu
Gly 945	Leu	Lys	Сув	Val	Ser 950	Phe	Leu	Asp	Gln	Ile 955	Ile	Pro	Thr	Ile	Leu 960
Asp	Val	Met	Arg	Thr 965	Сув	Ser	Gln	Ser	Leu 970	Leu	Glu	Phe	Tyr	Phe 975	Gln
Gln	Leu	Cys	Ser 980	Leu	Ile	Ile		Val 985	Arg	Gln	His	Ile	Arg 990	Pro	His
		995			Gln :		1000					1005	i		
Leu	Gln 1010	Ile	Thr	Leu	Val :	Ser 1015	Val	Ile	Glu	Ala	Ile 1020		Lys	Ala	Leu
Glu 1025	Gly	Glu	Phe	Lys	Arg 1	Leu	Val	Pro	Leu	Thr 1035		Thr	Leu	Phe	Leu 1040

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Val Val	Ile	Let	ı Glı	ı Ası	. n.	_									
Val				10	15 15	э Ьу	s Se	: Se	r As 10	р <b>Ly</b> 50	s Va	l Let	ı Se:	10!	
	Leu	Arg	106	ı Let	ı Glu	ı Se	r Phe	Gl 10	y Pr 65	o As	n Lei	ı Glu	1 Gl		Ser
His	Leu	Ile 107	: Th:	r Pro	Lye	s Ile	e Val	G1:	n Me	t Ala	a Glu	1 Phe		. Ser	Gly
Asn	Leu 1090	Gln	Arg	g Sei	Ala	11e	e Ile 95	Th:	r Il	e Gly	7 Lys		a Ala	a Lys	Asp
Val . 1105	Авр	Leu	Phe	e Glu	Met 111	Sei	Ser	Ar	g Il	e Val		Ser	Let	ı Lev	Arg 1120
Val :	Leu	Ser	Ser	Thr 112	Thr	Ser	Asp	Gl	Lei 11:	ı Sei 30	Lys	. Val	. Ile	Met 113	
Thr 1	Leu	Ser	Leu 114	Leu 0	Leu	Ile	e Gln	Met	: Gly 15	y Thr	Ser	Phe	Ala 115		Phe
Ile 1	Pro	Val 115	Ile 5	Asn	Glu	Val	. Leu 116	Met 0	: Lys	s Lys	His	Ile		His	Thr
Ile :	<b>Ty</b> r 1170	Asp	Asp	Leu	Thr	Asn 117	Arg	Ile	e Let	a Asn	Asn 118		Val	Leu	Pro
Thr I	Lys	Ile	Leu	Glu	Ala 119	Asn 0	Thr	Thr	'Ası	Tyr 119	Lys 5	Pro	Ala	Glu	Gln 1200
Met (	Glu	Ala	Ala	Asp 120	Ala 5	Gly	Val	Ala	Lys 121		Pro	Ile	Asn	Gln 121	
Val I	Leu	Lys	Ser 122	Ala 0	Trp	Asn	Ser	Ser 122	Gln 5	Gln	Arg	Thr	L <b>y</b> s 123		Asp
Trp G	ln	Glu 1235	Trp	Ser	Lys	Arg	Leu 124	Ser	Ile	Gln	Leu	Leu 124		Glu	Ser
Pro S	Ser 1250	His	Ala	Leu	Arg	Ala 125	Cys 5	Ser	Asn	Leu	Ala 126		Met	Tyr	Tyr
Pro L 1265	eu .	Ala	Lys	Glu	Leu 1270	Phe	Asn	Thr	Ala	Phe 127		Cys	Val	Trp	Thr 1280
Glu L	eu '	Tyr	Ser	Gln 128	Tyr 5	Gln	Glu	Asp	Leu 129		Gly	Ser	Leu	Cys 1295	
Ala L	eu :	Ser	Ser 1300	Pro	Leu	Asn	Pro	Pro 130	Glu 5	Ile	His	Gln	Thr 1310		Leu
Asn L	eu Y	Val 1315	Glu	Phe	Met	Glu	His 1320	Asp	Asp	Lys	Ala	Leu 1325		Ile	Pro
Thr G	ln 8 330	Ser	Leu	Gly	Glu	Tyr 1335	Ala	Glu	Arg	Сув	His 1340	Ala	Tyr	Ala	Lys
Ala L 1345	eu F	lis	Tyr	Lys	Glu 1350	Ile	Lys	Phe	Ile	Lys 1355	Glu	Pro	Glu	Asn	Ser 1360
Thr I	le 0	lu	Ser	Leu 1365	Ile	Ser	Ile	Asn	Asn 1370	Gln	Leu	Asn	Gln	Thr 1375	
Ala A	la 1	le	Gly 1380	Ile	Leu	Lys	His	Ala 1385	Gln	Gln	His		Ser 1390		Gln
Leu Ly	ув G 1	lu ' .395	Thr	Trp	Phe	Glu	L <b>y</b> s 1400	Leu	Glu	Arg	Trp	Glu 1405		Ala	Leu
His Al	la 1 410	yr i	Asn	Glu	Arg	Glu 1415	Lys	Ala	Gly	Asp	Thr 1420	Ser	Val	Ser	Val
Thr Le 1425	eu G	ly 1	Lys	Met	Arg 1430	Ser	Leu	His	Ala	Leu 1435		Glu '	Trp		Gln 1440
ieu Se	er G	ln 1	Leu :	Ala 1445	Ala .	Arg	Lys '	Trp	Lys 1450		Ser	Lys :		Gln 1455	Thr

45

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											-	-cor	ntir	ued	
Lys	Lys	Lev	11∈ 146	Ala	Pro	Leu	ı Ala	Ala 146	Gly	Ala	Arg	Trp	Gly 147	Leu '0	Gly
Glu	Trp	Asp 147	Met 5	: Leu	Glu	Gln	148	11e	. Ser	. Val	Met	Lys 148		Lys	Ser
Pro	Asp 149	Lys 0	Glu	Phe	Phe	Asp 149	Ala 5	Ile	Leu	Tyr	Leu 150		Lys	Asn	Asp
<b>Tyr</b> 150	Asp	Asn	Ala	Ser	Lys 151	His O	Ile	Leu	Asn	Ala 151		Asp	Leu	Leu	Val
Thr	Glu	Ile	Ser	Ala 152	Leu 5	Ile	Asn	Glu	Ser 153	Tyr 0	Asn	Arg	Ala	Tyr 153	Ser 5
Val	Ile	Val	Arg 154	Thr 0	Gln	Ile	Ile	Thr 154	Glu 5	Phe	Glu	Glu	Ile 155		Lys
Tyr	Lys	Gln 155		Pro	Pro	Asn	Ser 156		Lys	Lys	Leu	His 156		Gln	Asn
Leu	Trp 157	Thr 0	Lys	Arg	Leu	Leu 157	Gly 5	Cys	Gln	Lys	Asn 158		Asp	Leu	Trp
Gln 158	Arg 5	Val	Leu	Arg	Val 159	Arg 0	Ser	Leu	Val	Ile 159	L <b>y</b> s 5	Pro	Lys	Gln	Asp 1600
Leu	Gln	Ile	Trp	Ile 160	Lys 5	Phe	Ala	Asn	Leu 161	Cys 0	Arg	Lys	Ser	Gly 161	Arg 5
Met	Arg	Leu	Ala 162		Lys	Ala	Leu	Asn 162		Leu	Leu	Glu	Gly 163		Asn
Asp	Pro	Ser 163	Leu 5	Pro	Asn	Thr	Val 1640		Ala	Pro	Pro	Pro 164		Val	Tyr
Ala	Gln 165		Lys	Tyr	Ile	Trp 165	Ala 5	Thr	Gly	Ala	Tyr 1660		Glu	Ala	Leu
Asn 166	His 5	Leu	Ile	Gly	Phe 167	Thr	Ser	Arg	Leu	Ala 167		Asp	Leu	Gly	Leu 1680
Asp	Pro	Asn	Asn	Met 1685	Ile 5	Ala	Gln	Ser	Val 1690		Leu	Ser	Ser	Ala 1695	
Thr	Ala	Pro	Туr 1700	Val	Glu	Glu	Tyr	Thr 1705		Leu	Leu	Ala	Arg 1710		Phe
Leu	Lys	Gln 1715	Gly	Glu	Trp	Arg	Ile 1720		Thr	Gln	Pro	Asn 1725		Arg	Asn
Thr	Asn 1730	Pro	Asp	Ala	Ile	Leu 1735	Gly	Ser	Tyr	Leu	Leu 1740	Ala	Thr	His	Phe
Asp 1745	Lys	Asn	Trp	Tyr	Lys 1750	Ala )	Trp	His	Asn	Trp 1755	Ala	Leu	Ala	Asn	Phe 1760
Glu	Val	Ile	Ser	Met 1765	Val	Gln	Glu	Glu	Thr 1770		Leu	Asn	Gly	Gly 1775	
Asn	Asp	Asp	Asp	Asp	Asp	Thr	Ala	Val	Asn	Asn	Asp	Asn	Val	Arg	Ile

		14	15				148	0				148	5		
Pro	Asp 149	L <b>y</b> s	s Glu	ı Phe	Phe	Asp 149	Ala 5	Ile	Leu	Tyr	Leu 150		Lys	Asn	Asp
Туг 150	Asp	Asr	Ala	a Ser	Lys 151	His 0	Ile	Leu	Asn	Ala 151		Asp	Leu	Leu	Val 152
Thr	Glu	Ile	Ser	152	Leu !5	Ile	Asn	Glu	Ser 153		Asn	Arg	Ala	Tyr 153	Ser 5
Val	Ile	Val	. Arg		Gln	Ile	Ile	Thr 154		Phe	Glu	Glu	Ile 155		Lys
Tyr	Lys	Gln 155	Leu 5	Pro	Pro	Asn	Ser 156		Lys	Lys	Leu	His 156		Gln	Asn
Leu	Trp 157	Thr 0	Lys	Arg	Leu	Leu 157	Gly 5	Cys	Gln	Lys	Asn 158		Asp	Leu	Trp
Gln 158	Arg 5	Val	Leu	Arg	Val 159	Arg 0	Ser	Leu	Val	Ile 159		Pro	Lys	Gln	Asp 1600
Leu	Gln	Ile	Trp	160	Lys 5	Phe	Ala	Asn	Leu 161		Arg	Lys	Ser	Gly 161	Arg 5
Met	Arg	Leu	Ala 162	Asn 0	Lys	Ala	Leu	Asn 162		Leu	Leu	Glu	Gly 163		Asn
Asp	Pro	Ser 163		Pro	Asn	Thr	Val 1640		Ala	Pro	Pro	Pro 164		Val	Tyr
Ala	Gln 165	Leu 0	Lys	Tyr	Ile	Trp 165	Ala 5	Thr	Gly	Ala	Tyr 1660		Glu	Ala	Leu
Asn 166	His 5	Leu	Ile	Gly	Phe 167		Ser	Arg	Leu	Ala 1675		Asp	Leu	Gly	Leu 1680
Asp	Pro	Asn	Asn	Met 168	Ile 5	Ala	Gln	Ser	Val 169	Lys 0	Leu	Ser	Ser	Ala 1695	
Thr	Ala	Pro	Tyr 170	Val 0	Glu	Glu	Tyr	Thr 1705	Lys S	Leu	Leu	Ala	Arg 1710		Phe
Leu	Lys	Gln 171	Gly 5	Glu	Trp	Arg	Ile 1720		Thr	Gln	Pro	Asn 1725		Arg	Asn
Thr	Asn 1730	Pro	Asp	Ala	Ile	Leu 1735	Gly	Ser	Tyr	Leu	Leu 1740		Thr	His	Phe
Asp 1745	L <b>y</b> s	Asn	Trp	Tyr	Lys 1750	Ala )	Trp	His	Asn	Trp 1755		Leu	Ala	Asn	Phe 1760
Glu	Val	Ile	Ser	Met 176	Val	Gln	Glu	Glu	Thr 1770	Lys )	Leu	Asn	Gly	Gly 1775	
Asn	Asp	Asp	Asp 1780	Asp 0	Asp	Thr	Ala	Val 1785		Asn	Asp	Asn	Val 1790		Ile
Asp	Gly	Ser 1795	Ile	Leu	Gly	Ser	Gl <b>y</b> 1800	Ser	Leu	Thr	Ile	Asn 1805		Asn	Arg
Tyr	Pro 1810		Glu	Leu	Ile	Gln 1815		His	Val	Val	Pro 1820		Ile	Lув	Gly
Phe 1825		His	Ser	Ile	Ser 1830		Leu	Glu	Thr	Ser 1835		Leu	Gln	Asp	Thr 1840
Leu	Arg	Leu	Leu	Thr 1845		Leu	Phe	Asn	Phe 1850	Gly	Gly	Ile	Lys	Glu 1855	
Ser	Gln	Ala	Met 1860		Glu	Gly		Asn 1865		Met	Lys		Glu 1870		Trp

Leu Glu Val Leu Pro Gln Leu Ile Ser Arg Ile His Gln Pro Asp Pro

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		-concinuea
1875	1880	1885
Thr Val Ser Asn Ser 1890	r Leu Leu Ser Leu L 1895	eu Ser Asp Leu Gly Lys Ala 1900
His Pro Gln Ala Leo 1905	ı Val Tyr Pro Leu T 1910	hr Val Ala Ile Lys Ser Glu 1915 1920
Ser Val Ser Arg Glr 192		er Ile Ile Glu Lys Ile Arg 930 1935
Ile His Ser Pro Val	Leu Val Asn Gln A 1945	la Glu Leu Val Ser His Glu 1950
Leu Ile Arg Val Ala 1955	val Leu Trp His G 1960	lu Leu Trp Tyr Glu Gly Leu 1965
Glu Asp Ala Arg Arg 1970	Gln Phe Phe Val Gl 1975	lu His Asn Ile Glu Lys Met 1980
Phe Ser Thr Leu Glu 1985	Pro Leu His Lys Hi 1990	is Leu Gly Asn Glu Pro Gln 1995 2000
Thr Leu Ser Glu Val 200	Ser Phe Gln Lys Se 5 20	er Phe Gly Arg Asp Leu Asn 2010 2015
Asp Ala Tyr Glu Trp 2020	Leu Asn Asn Tyr Ly 2025	ys Lys Ser Lys Asp Ile Asn 2030
2035	2040	or Asn Val Phe Arg Lys Ile 2045
Thr Arg Gln Ile Pro 2050	Gln Leu Gln Thr Le 2055	eu Asp Leu Gln His Val Ser 2060
Pro Gln Leu Leu Ala 2065	Thr His Asp Leu Gl 2070	u Leu Ala Val Pro Gly Thr 2075 2080
208	5 20	e Ala Lys Phe Glu Pro Leu 90 2095
Phe Ser Val Ile Ser 2100	Ser Lys Gln Arg Pr 2105	o Arg Lys Phe Ser Ile Lys 2110
2115	2120	l Leu Lys Gly His Glu Asp 2125
2130	2135	u Phe Gly Leu Val Asn Thr 2140
2145	2150	s Arg His Leu Asp Ile Gln 2155 2160
2165	21	
2180	2185	u Ile Arg Glu His Arg Asp 2190
2195	2200	n Trp Val Met Leu Gln Met 2205
2210	2215	u Gln Lys Ile Glu Val Phe 2220
Thr Tyr Ala Leu Asp 2225	Asn Thr Lys Gly Gli 2230	n Asp Leu Tyr Lys Ile Leu 2235 2240
2245	225	
2260	2265	t Thr Gly Tyr Ile Leu Gly 2270
2275	2280	t Leu Asp Arg Ile Thr Gly 2285
Lys Val Ile His Ile 2290	Asp Phe Gly Asp Cys 2295	s Phe Glu Ala Ala Ile Leu 2300

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Arg Glu Lys Tyr Pro Glu Lys Val Pro Phe Arg Leu Thr Arg Met Leu 2305 2310 2315

Thr Tyr Ala Met Glu Val Ser Gly Ile Glu Gly Ser Phe Arg Ile Thr 2325 2330 2330 2335

Cys Glu Asn Val Met Arg Val Leu Arg Asp Asn Lys Glu Ser Leu Met 2340 2345 2350

Ala Ile Leu Glu Ala Phe Ala Leu Asp Pro Leu Ile His Trp Gly Phe 2355 2360 2365

Asp Leu Pro Pro Gln Lys Leu Thr Glu Gln Thr Gly Ile Pro Leu Pro 2370 2375 2380

Leu Ile Asn Pro Ser Glu Leu Leu Arg Lys Gly Ala Ile Thr Val Glu 2385 2390 2395 2400

Glu Ala Ala Asn Met Glu Ala Glu Gln Gln Asn Glu Thr Arg Asn Ala 2405 2410 2415

Arg Ala Met Leu Val Leu Arg Arg Ile Thr Asp Lys Leu Thr Gly Asn  $2420 \hspace{1.5cm} 2425 \hspace{1.5cm} 2430 \hspace{1.5cm}$ 

Asp Ile Lys Arg Phe Asn Glu Leu Asp Val Pro Glu Gln Val Asp Lys 2435  $\phantom{0}2440$   $\phantom{0}2445$ 

Leu Ile Gln Gln Ala Thr Ser Ile Glu Arg Leu Cys Gln His Tyr Ile 2450 2455 2460

Gly Trp Cys Pro Phe Trp 2465 2470

- (2) INFORMATION FOR SEQ ID NO:4:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 2474 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Saccharomyces cerevisiae
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Asn Lys Tyr Ile Asn Lys Tyr Thr Thr Pro Pro Asn Leu Leu Ser 1  $\phantom{\bigg|}$  10  $\phantom{\bigg|}$  15

Leu Arg Gln Arg Ala Glu Gly Lys His Arg Thr Arg Lys Lys Leu Thr 20 25 30

His Lys Ser His Ser His Asp Asp Glu Met Ser Thr Thr Ser Asn Thr 35 40 45

Asp Ser Asn His Asn Gly Pro Asn Asp Ser Gly Arg Val Ile Thr Gly 50

Ser Ala Gly His Ile Gly Lys Ile Ser Phe Val Asp Ser Glu Leu Asp 65 70 75 80

Thr Thr Phe Ser Thr Leu Asn Leu Ile Phe Asp Lys Leu Lys Ser Asp 85 90 95

Val Pro Gln Glu Arg Ala Ser Gly Ala Asn Glu Leu Ser Thr Thr Leu  $100 \hspace{1.5cm} 105 \hspace{1.5cm} 110 \hspace{1.5cm}$ 

Thr Ser Leu Ala Arg Glu Val Ser Ala Glu Gln Phe Gln Arg Phe Ser 115 120 125

Asn Ser Leu Asn Asn Lys Ile Phe Glu Leu Ile His Gly Phe Thr Ser 130 140

Ser Glu Lys Ile Gly Gly Ile Leu Ala Val Asp Thr Leu Ile Ser Phe 145 150 155 160

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Tyr	Leu	Ser	Thr	Glu 165		Leu	Pro	Asn	Gln 170		Ser	Arg	Leu	Ala 175	Asn
Tyr	Leu	Arg	Val 180	Leu	Ile	Pro	Ser	Ser 185		Ile	Glu	Val	Met 190		Leu
Ala	Ala	Asn 195		Leu	Gly	Arg	Leu 200	Thr	Val	Pro	Gly	Gly 205	Thr	Leu	Thr
Ser	Asp 210	Phe	Val	Glu	Phe	Glu 215		Arg	Thr	Сув	Ile 220	Asp	Trp	Leu	Thr
Leu 225	Thr	Ala	Asp	Asn	Asn 230	Ser	Ser	Ser	Ser	Lys 235	Leu	Glu	Tyr	Arg	Arg 240
His	Ala	Ala	Leu	Leu 245	Ile	Ile	Lys	Ala	Leu 250	Ala	Asp	Asn	Ser	Pro 255	
Leu	Leu	Tyr	Pro 260	Tyr	Val	Asn	Ser	Ile 265	Leu	Asp	Asn	Ile	Trp 270	Val	Pro
Leu	Arg	Авр 275	Ala	Lys	Leu	Ile	11e 280	Arg	Leu	Asp	Ala	Ala 285	Val	Ala	Leu
Gly	Lys 290	Cys	Leu	Thr	Ile	Ile 295	Gln	Asp	Arg	Asp	Pro 300	Ala	Leu	Gly	Lys
Gln 305	Trp	Phe	Gln	Arg	Leu 310	Phe	Gln	Gly	Суѕ	Thr 315	His	Gly	Leu	Ser	Leu 320
Asn	Thr	Asn	Asp	Ser 325	Val	His	Ala	Thr	Leu 330	Leu	Val	Phe	Arg	Glu 335	Leu
Leu	Ser	Leu	Lys 340	Ala	Pro	Tyr	Leu	Arg 345	Asp	Lys	Tyr	Asp	Asp 350	Ile	Tyr
Lys	Ser	Thr 355	Met	Lys	Tyr	Lys	Glu 360	Tyr	Lys	Phe	Asp	Val 365	Ile	Arg	Arg
Glu	Val 370	Tyr	Ala	Ile	Leu	Pro 375	Leu	Leu	Ala	Ala	Phe 380	qaA	Pro	Ala	Ile
Phe 385	Thr	Lys	Lys	Tyr	Leu 390	Asp	Arg	Ile	Met	Val 395	His	Tyr	Leu	Arg	Tyr 400
			Ile	405					410			_	-	415	
			Ser 420					425					430		
		435	Met				440					445			
Thr	Lys 450	Phe	Lys	Val	Arg	<b>Lys</b> 455	Gln	Phe	Glu	Lys	Asp 460	Leu	Phe	Tyr	Cys
Ile 465	Gly	Lys	Leu	Ala	Cys 470	Ala	Leu	Gly	Pro	Ala 475	Phe	Ala	Lys	His	Leu 480
Asn	Lys	Asp	Leu	Leu 485	Asn	Leu	Met	Leu	Asn 490	Cys	Pro	Met	Ser	Asp 495	His
Met			500					505					510		
Ser		515					520					525			
	530					535			-	•	540				
Ser 545					550			-		555				-	560
Thr				565					570					575	
Cys	Phe	Lys	Met	Leu	Gln	Leu	Ile	His	His	Gln	Tyr	Ser	Leu	Thr	Glu

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			580	)				585	5				596	)	
Phe	va]	595	Lei	ı Ile	∍ Thr	: Ile	60¢		rIle	e Glu	ı His	605		Sei	: Ser
Val	Arg 610	J Lys	s Leu	ı Ala	Ala	Lev 615	Th:	Ser	с Сув	a Asp	620		e Ile	e Lys	Asp
Asp 625	Ile	e Cys	s Lys	Gln	Thr 630	Ser	· Vai	l His	Ala	635		Ser	Va]	Ser	Glu 640
Val	Leu	Se:	Lys	645	Leu	Met	: Ile	e Ala	11e		Asp	Pro	Va]	Ala 655	Glu
Ile	Arg	Leu	660	Ile	Leu	Glm	Hi:	665	Gly	7 Ser	Asn	Phe	Asp 670		Gln
Leu	Ala	Glr 675	Pro	qaA q	Asn	Leu	Arg 680	Leu	Leu	Phe	Met	Ala 685		Asn	Asp
Glu	Ile 690	Phe	Gly	Ile	Gln	Leu 695	Glu	Ala	Ile	Lys	700	Ile	Gly	Arg	Leu
Ser 705	Ser	Val	Asn	Pro	Ala 710	Tyr	Val	. Val	Pro	Ser 715		Arg	Lys	Thr	Leu 720
Leu	Glu	Leu	Leu	Thr 725	Gln	Leu	Lys	Phe	Ser 730		Met	Pro	Lys	<b>Lys</b> 735	Lys
Glu	Glu	Ser	Ala 740	Thr	Leu	Leu	Сув	Thr 745	Leu	Ile	Asn	Ser	Ser 750	Asp	Glu
Val	Ala	Lys 755	Pro	Tyr	Ile	Asp	Pro 760	Ile	Leu	Asp	Val	Ile 765	Leu	Pro	L <b>y</b> s
Сув	Gln 770	Asp	Ala	Ser	Ser	Ala 775	Val	Ala	Ser	Thr	Ala 780	Leu	Lys	Val	Leu
Gly 785	Glu	Leu	Ser	Val	Val 790	Gly	Gly	Lys	Glu	Met 795	Thr	Arg	Tyr	Leu	<b>Lys</b> 800
Glu	Leu	Met	Pro	Leu 805	Ile	Ile	Asn	Thr	Phe 810	Gln	Asp	Gln	Ser	Asn 815	Ser
Phe	Lys	Arg	Asp 820	Ala	Ala	Leu	Thr	Thr 825	Leu	Gly	Gln	Leu	Ala 830	Ala	Ser
Ser	Gly	Tyr 835	Val	Val	Gly	Pro	Leu 840	Leu	Asp	Tyr	Pro	Glu 845	Leu	Leu	Gly
	850					855				Asn	860				
865					870					Ala 875					880
His	Arg	Glu	Ile	Glu 885	Val	Thr	Ser	naA	Ser 890	Lys	Ser	Ser	Val	Glu 895	Gln
Asn	Ala	Pro	Ser 900	Ile	Asp	Ile		Leu 905		Met	Gln	Gly	Val 910	Ser	Pro
Ser	Asn	Asp 915	Glu	Tyr	Tyr	Pro	Thr 920	Val	Val	Ile	His	Asn 925	Leu	Met	Lys
Ile	Leu 930	Asn	Asp	Pro		Leu 935	Ser	Ile	His	His	Thr 940	Ala	Ala	Ile	Gln
Ala 945	Ile	Met	His	Ile	Phe 950	Gln	Asn	Leu	Gly	Leu 955	Arg	Cys	Val	Ser	Phe 960
Leu	Asp	Gln	Ile	Ile 965	Pro	Gly	Ile	Ile	Leu 970	Val	Met	Arg	Ser	Сув 975	Pro
Pro	Ser	Gln	Leu 980	Asp	Phe	Tyr	Phe	Gln 985	Gln	Leu	Gly		Leu 990	Ile	Ser
Ile	Val	Lys 995	Gln	His	Ile	Arg	Pro 1000	His	Val	Glu		Ile 1005		Gly	Val

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Ile Arg Glu Pho	e Phe Pro	Ile Ile 1015	e Lys Le	u Gln Ile 102	Thr Ile	e Ile Ser
Val Ile Glu Ser 1025	r Ile Ser 103	Lys Ala	Leu Gl	u Gly Glu 1035	Phe Lys	Arg Phe
Val Pro Glu Thi	Leu Thr 1045	Phe Phe	e Leu As 10	p Ile Leu 50	Glu Asn	Asp Gln 1055
Ser Asn Lys Arc	J lle Val	Pro Ile	Arg Ile 1065	e Leu Lys	Ser Leu 107	
Phe Gly Pro Asr 1075	ı Leu Glu	Asp Tyr	Ser Hi	s Leu Ile	Met Pro	Ile Val
Val Arg Met Thr 1090	Glu Tyr	Ser Ala 1095	Gly Ser	Leu Lys 110		Ser Ile
Ile Thr Leu Gly 1105	Arg Leu 1110	Ala Lys	Asn Ile	e Asn Leu 1115	Ser Glu	Met Ser 1120
Ser Arg Ile Val	Gln Ala 1125	Leu Val	Arg Ile		Asn Gly	Asp Arg 1135
Glu Leu Thr Lys	Ala Thr	Met Asn	Thr Leu 1145	ser Leu	Leu Leu 115	
Leu Gly Thr Asp 1155	Phe Val	Val Phe 116	Val Pro	Val Ile	Asn Lys 1165	Ala Leu
Leu Arg Asn Arg 1170	Ile Gln	His Ser 1175	Val Tyr	Asp Gln 118		Asn Lys
Leu Leu Asn Asn 1185	Glu Cys 1190	Leu Pro	Thr Asn	Ile Ile 1195	Phe Asp	Lys Glu 1200
Asn Glu Val Pro	Glu Arg 1205	Lys Asn	Tyr Glu 121		Met Gln	Val Thr 1215
Lys Leu Pro Val 122	Asn Gln O	Asn Ile	Leu Lys 1225	Asn Ala	Trp Tyr 1230	
Gln Gln Lys Thr 1235	Lys Glu	Asp Trp 124	Gln Glu 0	Trp Ile	Arg Arg 1245	Leu Ser
Ile Gln Leu Leu 1250	Lys Glu	Ser Pro 1255	Ser Ala	Cys Leu 1260		Cys Ser
Ser Leu Val Ser 1265	Val Tyr 1270	Tyr Pro	Leu Ala	Arg Glu 1275	Leu Phe	Asn Ala 1280
Ser Phe Ser Ser	Cys Trp 1285	Val Glu	Leu Gln 129	Thr Ser	Tyr Gln	Glu Asp 1295
Leu Ile Gln Ala 1300	Leu Cys	Lys Ala	Leu Ser 1305	Ser Ser	Glu Asn 1310	
Glu Ile Tyr Gln 1315	Met Leu	Leu Asn 1320		Glu Phe	Met Glu 1325	His Asp
Asp Lys Pro Leu 1330	Pro Ile	Pro Ile 1335	His Thr	Leu Gly 1340	Lys Tyr	Ala Gln
Lys Cys His Ala 1345	Phe Ala 1 1350	Lys Ala	Leu His	Tyr Lys 1355	Glu Val	Glu Phe 1360
Leu Glu Glu Pro	Lys Asn : 1365	Ser Thr	Ile Glu 1370	Ala Leu )	Ile Ser	Ile Asn 1375
Asn Gln Leu His 1380	Gln Thr	Asp Ser	Ala Ile 1385	Gly Ile	Leu Lys 1390	
Gln Gln His Asn 1395	Glu Leu (	Gln Leu 1400	Lys Glu		Tyr Glu 1405	Lys Leu
Gln Arg Trp Glu 1410	Asp Ala I	Ceu Ala 1415	Ala Tyr	Asn Glu 1420	Lys Glu	Ala Ala

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_															
Gly 142	Glu 5	ı Asp	) Se	r Val	143	ı Val	Met	. Met	Gly	Lys 143	Leu 5	Arg	g Sei	r Le	1 Tyr 1440
Ala	Lev	ı Gly	/ Gl	ı Trg	Glu 15	ı Glu	ı Let	ser	Lys 145	Leu 10	Ala	Sei	c Glu	1 Lys	Trp
Gly	Thr	Ala	146	s Pro	Glu	Val	. Lys	Lys 146		Met	Ala	Pro	Let 147		a Ala
Gly	Ala	Ala 147	Trp	o Gly	Leu	Glu	Gln 148	Trp	Asp	Glu	Ile	: Ala		тул	Thr
Ser	Val 149	. Met	: Lys	s Ser	Gln	Ser	Pro	Asp	Lys	Glu	Phe 150	Tyr 0	: Asp	Ala	lle
Leu 150	С <b>у</b> в 5	Leu	His	Arg	Asn 151	Asn 0	Phe	Lys	Lys	Ala 151		Val	. His	Il∈	Phe 1520
Asn	Ala	Arg	Asp	Leu 152	Leu 5	Val	Thr	Glu	Leu 153		Ala	Leu	Val	Asn 153	Glu 5
Ser	Tyr	Asn	Arg 154	Ala 0	Tyr	Asn	Val	Val 154		Arg	Ala	Gln	Ile 155		: Ala
Glu	Leu	Glu 155	Glu 5	ılle	Ile	Lys	Tyr 156	L <b>y</b> s	Lys	Leu	Pro	Gln 156		Ser	Asp
Lys	Arg 157	Leu 0	Thr	Met	Arg	Glu 157	Thr 5	Trp	Asn	Thr	Arg 158	Leu 0	Leu	Gly	Cys
Gln 1585	Lys 5	Asn	Ile	asp	Val	Trp 0	Gln	Arg	Ile	Leu 159		Val	Arg	Ser	Leu 1600
Val	Ile	Lys	Pro	L <b>y</b> s 160		Asp	Ala	Gln	Val		Ile	Lys	Phe	Ala 161	
Leu	Cys	Arg	<b>Lys</b> 162	Ser 0	Gly	Arg	Met	Ala 1625	Leu	Ala	Lys	Lys	Val 163		Asn
Thr	Leu	Leu 163	Glu 5	Glu	Thr	qaA	Asp 1646	Pro	Авр	His	Pro	Asn 164		Ala	Lys
Ala	Ser 1650	Pro	Pro	Val	Val	Tyr 1655	Ala	Gln	Leu	Lys	Tyr 1660		Trp	Ala	Thr
Gly 1665	Leu	Gln	Asp	Glu	Ala 1670	Leu )	Lys	Gln	Leu	Ile 1675	Asn	Phe	Thr	Ser	Arg 1680
Met	Ala	His	Asp	Leu 1689	Gly	Leu	qaA	Pro	Asn 1690		Met	Ile	Ala	Gln 169	
Val	Pro	Gln	Gln 170	Ser	Lys	Arg	Val	Pro 1705		His	Val	Glu	Asp 1710		Thr
Lys	Leu	Leu 1715	Ala	Arg	Сув	Phe	Leu 1720	Lys )	Gln	Gly	Glu	Trp 1725		Val	Cys
Leu	Gln 1730	Pro	Lys	Trp	Arg	Leu 1735	Ser	Asn	Pro	Asp	Ser 1740		Leu	Gly	Ser
Tyr 1745	Leu	Leu	Ala	Thr	His 1750	Phe	Asp	Asn	Thr	Trp 1755	Tyr	Lys	Ala	Trp	His 1760
Asn '	Trp	Ala	Leu	Ala 1765	Asn	Phe	Glu	Val	Ile 1770	Ser	Met	Leu	Thr	Ser 1775	
Ser :	Lys	Lys	Lys 1780	Gln	Glu	Gly	Ser	Asp 1785	Ala	Ser	Ser	Val	Thr 1790		Ile
Asn (	Glu	Phe 1795	Asp	Asn	Gly	Met	Ile 1800	Gly	Val.	Asn		Phe 1805		Ala	Lys
Glu Y	Val 1810	His	Tyr	Ser	Ser .	Asn 1815	Leu	Ile	His .		His 1820		Ile	Pro	Ala
[le ] [825	L <b>y</b> s	Gly	Phe		His 1830		Ile	Ser 1		Ser (	Glu	Ser	Ser	Ser	Leu 1840
3ln <i>1</i>	Asp .	Ala	Leu	Arg	Leu :	Leu '	Thr	Leu '	rp :	Phe '	Thr :	Phe	Gly	Gly	Ile

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					184	5				185	0				185	5
:	Pro	Glu	Ala	Thr 186		Ala	Met	His	Glu 186		Phe	Asn	Leu	Ile 187		Ile
•	Gly	Thr	Trp 187		Glu	Val	Leu	Pro 188		Leu	Ile	Ser	Arg 188		His	Gln
1	Pro	Asn 189		Ile	Val	Ser	Arg 189		Leu	Leu	Ser	Leu 190		Ser	Asp	Leu
	3ly 190		Ala	His	Pro	Gln 191		Leu	Val	Tyr	Pro 191		Met	Val	Ala	Ile 1920
]	Lуs	Ser	Glu	Ser	Leu 192		Arg	Gln	Lys	Ala 193	Ala 0	Leu	Ser	Ile	Ile 193	
]	Lys	Met	Arg	Ile 194		Ser	Pro	Val	Leu 194		Asp	Gln	Ala	Glu 195		Val
:	Ser	His	Glu 195		Ile	Arg	Met	Ala 196		Leu	Trp	His	Glu 196		Trp	Tyr
(	3lu	Gly 197		Asp	Asp	Ala	Ser 197		Gln	Phe	Phe	Gly 1980		His	Asn	Thr
	31u 198	Lys 5	Met	Phe	Ala	Ala 199		Glu	Pro	Leu	<b>Tyr</b> 199		Met	Leu	Lys	Arg 2000
(	3ly	Pro	Glu	Thr	Leu 200		Glu	Ile	Ser	Phe 201	Gln 0	Asn	Ser	Phe	Gly 2015	
1	Asp	Leu	Asn	Asp 202		Tyr	Glu	Trp	Leu 202		Asn	Tyr	Lys	Lys 2030		Lys
1	4sp	Val	Ser 203		Leu	Asn	Gln	Ala 2040		Asp	Ile	Tyr	Tyr 204		Val	Phe
1	arg	Lys 2050	Ile	Gly	Lys	Gln	Leu 2055		Gln	Leu	Gln	Thr 2060	Leu )	Glu	Leu	Gln
	lis 2065		Ser	Pro	Lys	Leu 207		Ser	Ala	His	Asp 2075		Glu	Leu	Ala	Val 2080
E	Pro	Gly	Thr	Arg	Ala 208	Ser 5	Gly	Gly	Lys	Pro 2090	Ile O	Val	Lys	Ile	Ser 2095	
F	he	Glu	Pro	Val 210		Ser	Val	Ile	Ser 2105		Lys	Gln	Arg	Pro 2110		Lys
E	he	Сув	Ile 211		Gly	Ser	Asp	Gly 2120		Asp	Tyr	Lys	Tyr 2125		Leu	Lys
G	Зly	His 2130		Asp	Ile	Arg	Gln 2135		Ser	Leu	Val	Met 2140		Leu	Phe	Gly
	eu 145		Asn	Thr	Leu	Leu 215(		Asn	Asp	Ala	Glu 2155		Phe	Arg	Arg	His 2160
Ι	eu	Asp	Ile	Gln	Gln 216		Pro	Ala	Ile	Pro 2170	Leu )	Ser	Pro	Lys	Ser 2175	-
I	eu	Leu	Gly	Trp 2180		Pro	Asn	Ser	Asp 2185		Phe	His	Val	Leu 2190		Arg
G	lu	His	Arg 2195		Ala	Lys		Ile 2200		Leu	Asn	Ile	Glu 2205		Trp	Val
M	let	Leu 2210		Met	Ala	Pro	Asp 2215		Asp	Asn	Leu	Thr 2220		Leu	Gln	Lys
	al 225		Val	Phe	Thr	Tyr 2230		Leu	Asn	Asn	Thr 2235		Gly	Gln	Asp	Leu 2240
Т	yr	Lys	Val	Leu	Trp 2245		Lys	Ser	Arg	Ser 2250	Ser	Glu	Thr	Trp	Leu 2255	
A	.rg	Arg	Thr	Thr 2260		Thr	Arg		Leu 2265		Val	Met	Ser	Met 2270		Gly

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Tyr Ile Leu Gly Leu Gly Asp Arg His Pro Ser Asn Leu Met Leu Asp 2275 2280 2285	
Arg Ile Thr Gly Lys Val Ile His Ile Asp Phe Gly Asp Cys Phe Glu 2290 2295 2300	
Ala Ala Ile Leu Arg Glu Lys Phe Pro Glu Lys Val Pro Phe Arg Leu 2305 2310 2315 2320	
Thr Arg Met Leu Thr Tyr Ala Met Glu Val Ser Gly Ile Glu Gly Ser 2325 2330 2335	
Phe Arg Ile Thr Cys Glu Asn Val Met Lys Val Leu Arg Asp Asn Lys 2340 2345 2350	
Gly Ser Leu Met Ala Ile Leu Glu Ala Phe Ala Phe Asp Pro Leu Ile 2355 2360 2365	
Asn Trp Gly Phe Asp Leu Pro Thr Lys Lys Ile Glu Glu Glu Thr Gly 2370 2375 2380	
Ile Gln Leu Pro Val Met Asn Ala Asn Glu Leu Leu Ser Asn Gly Ala 2385 2390 2395 2400	
Ile Thr Glu Glu Glu Val Gln Arg Val Glu Asn Glu His Lys Asn Ala 2405 2410 2415	
Ile Arg Asn Ala Arg Ala Met Leu Val Leu Lys Arg Ile Thr Asp Lys 2420 2425 2430	
Leu Thr Gly Asn Asp Ile Arg Arg Phe Asn Asp Leu Asp Val Pro Glu 2435 2440 2445	
Gln Val Asp Lys Leu Ile Gln Gln Ala Thr Ser Val Glu Asn Leu Cys 2450 2455 2460	
Gln His Tyr Ile Gly Trp Cys Pro Phe Trp 2465 2470	
(2) INFORMATION FOR SEQ ID NO:5:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 64 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:	
CCGGATCCCG TCGAGCTTCA GTTGAACTAC GGCGTGCTTC TGTAGCCATG GGAGTGCAGG	60
TGGA	64
(2) INFORMATION FOR SEQ ID NO:6:	
(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 28 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:	
GGCCGGAATT CTCATTCCAG TTTTAGAA	28

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
  (A) LENGTH: 7 amino acids
  (B) TYPE: amino acid
  (D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: DNA (genomic)
     (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:
 Thr Tyr Asp Pro Asn Gln Pro
 (2) INFORMATION FOR SEQ ID NO:8:
       (i) SEQUENCE CHARACTERISTICS:
            (A) LENGTH: 6 amino acids
(B) TYPE: amino acid
            (D) TOPOLOGY: linear
     (ii) MOLECULE TYPE: peptide
     (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:
His Ile Asp Phe Gly Asp
1 5
 (2) INFORMATION FOR SEQ ID NO:9:
       (i) SEQUENCE CHARACTERISTICS:
            (A) LENGTH: 6 amino acids
            (B) TYPE: amino acid
(D) TOPOLOGY: linear
     (ii) MOLECULE TYPE: peptide
     (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:
Asn Asp Gln Val Phe Glu
(2) INFORMATION FOR SEQ ID NO:10:
      (i) SEQUENCE CHARACTERISTICS:
            (A) LENGTH: 18 base pairs(B) TYPE: nucleic acid
            (C) STRANDEDNESS: single
            (D) TOPOLOGY: linear
     (ii) MOLECULE TYPE: cDNA
     (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:
GAGCCACCAC GATTTGCT
(2) INFORMATION FOR SEQ ID NO:11:
      (i) SEQUENCE CHARACTERISTICS:
           (A) LENGTH: 64 base pairs
            (B) TYPE: nucleic acid
            (C) STRANDEDNESS: single
           (D) TOPOLOGY: linear
    (ii) MOLECULE TYPE: cDNA
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
CCGGATCCCG TCGAGCTTCA GTTGAACTAC GGCGTGCTTC TGTAGCCATG GCGGCGGCCG
                                                                              60
TTCC
                                                                              64
(2) INFORMATION FOR SEQ ID NO:12:
     (i) SEQUENCE CHARACTERISTICS:
           (A) LENGTH: 28 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single
           (D) TOPOLOGY: linear
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(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:	
GGCCGGAATT CTCAATCAAT ATCCACTA	28
(2) INFORMATION FOR SEQ ID NO:13:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 28 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(ii) MOLECULE TYPE: cDNA	
(iii) HYPOTHETICAL: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:	
GGGGGATCCA CNTAYGAYCC NAAYCARC	28
(2) INFORMATION FOR SEQ ID NO:14:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 27 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:	
GCGGAATTCR TCNCCRAART CDATRTG	27
(2) INFORMATION FOR SEQ ID NO:15:	
(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 26 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:	

What is claimed is:

GGGGGATCCA AYGAYCARGT NTTYGA

1. An isolated, purified cDNA molecule which encodes RAFT1, a protein having the amino acid sequence as shown in SEQ ID NO:2 wherein the acronym RAFT connotes a rapamycin and FKBP12 target.

2. The isolated, purified cDNA molecule of claim 1 which comprises the nucleotide sequence as shown in SEQ ID NO:1, nucleotides 64-7710.

- 3. An isolated, purified intron-free DNA molecule con- 55 sisting of at least 20 contiguous nucleotides encoding all or a portion of the amino acid sequence as shown in SEQ ID NO: 2.
- 4. An isolated, purified intron-free DNA molecule consisting of at least 20 contiguous nucleotides of the sequence 60 as shown in SEQ ID NO: 1.
- 5. An isolated DNA molecule encoding a rat RAFT protein obtained by a method comprising the steps of:
  - (a) probing a library of rat cDNA sequences with a probe selected from the sequence shown in SEQ ID NO: 1;

(b) isolating a rat cDNA molecule which (i) hybridizes to the probe, (ii) contains a complete open reading frame encoding a polypeptide of about 2550 amino acids, and (iii) encodes a rat RAFT protein,

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wherein said rat RAFT protein binds to FKBP12 in the presence of 1 to 10 nM rapamycin but not in the absence of 1 to 10 nM rapamycin.

- 6. An isolated DNA molecule encoding a rat RAFT protein obtained by a method comprising the steps of:
  - (a) amplifying a DNA sequence using (i) at least one primer which comprises at least 10 contiguous nucleotides selected from the sequence shown in SEQ ID NO: 1 and (ii) a template which comprises rat cDNA or mRNA; and
  - (b) isolating an amplified DNA sequence which contains a complete open reading frame encoding a polypeptide of about 2550 amino acids encoding a rat RAFT protein,
- which comprises at least 15 contiguous nucleotides 65 wherein said rat RAFT protein binds to FKBP12 in the presence of 1 to 10 nM rapamycin but not in the absence of 1 to 10 nM rapamycin.

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- 7. An isolated DNA molecule encoding a rat RAFT protein identified by a process comprising the steps of:
  - (a) annealing a set of mixed oligonucleotides to a rat cDNA library, each member of said set of mixed oligonucleotides encoding a sequence of at least six 5 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:2; and
  - (b) isolating a rat cDNA molecule which (i) anneals to at least one member of the set of mixed oligonucleotides, (ii) contains a complete open reading frame encoding a polypeptide of about 2550 amino acids, and (iii) encodes a rat RAFT protein,

wherein said RAFT protein binds to FKBP12 in the presence of 1 to 10 nM rapamycin but not in the absence of 1 to 10 nM rapamycin.

- 8. An isolated DNA molecule encoding a rat RAFT protein according to claim 7, wherein two sets of mixed oligonucleotides are annealed.
- 9. An isolated DNA molecule having a nucleotide sequence, or a degenerate sequence thereof, obtained by a 20 method committee the sequence and 20 method committee the seq method comprising the steps of:
  - (a) probing a library of rat cDNA molecules with a probe which comprises at least 15 contiguous nucleotides
  - (b) isolating a rat cDNA molecule which (i) hybridizes to the probe, (ii) contains a complete open reading frame

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encoding a polypeptide of about 2550 amino acids, and (iii) encodes a rat RAFT protein,

wherein said RAFT protein binds to FKBP12 in the presence of 1 to 10 nM rapamycin but not in the absence of 1 to 10 nM rapamycin.

- 10. A method of isolating a DNA molecule encoding a mammalian RAFT protein comprising the steps of:
- (a) probing a library of rat cDNA sequences with a probe which comprises at least 15 contiguous nucleotides selected from the sequence shown in SEQ ID NO: 1;
- (b) isolating a rat cDNA molecule which (i) hybridizes to the probe, (ii) contains a complete open reading frame encoding a polypeptide of about 2550 amino acids, and (iii) encodes a rat RAFT protein,

wherein said rat RAFT protein binds to FKBP12 in the presence of 1 to 10 nM rapamycin but not in the absence of 1 to 10 nM rapamycin.

- 11. The method of claim 10 wherein the probe comprises at least 20 contiguous nucleotides encoding all or a portion of the amino acid sequence as shown in SEQ ID NO:2.
- 12. The method of claim 10 wherein the probe comprises selected from the sequence shown in SEQ ID NO: 1; 25 at least 20 contiguous nucleotides as shown in SEQ ID

# UNITED STATES PATENT AND TRADEMARK OFFICE **CERTIFICATE OF CORRECTION**

PATENT NO.

: 6,492,106 B1

Page 1 of 1

APPLICATION NO.: 08/305790

DATED

: December 10, 2002

INVENTOR(S)

: David M. Sabatini et al.

It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

On Title Page, should read, under (73) Assignee:

--Sloan-Kettering Institute for Cancer Research, New York, NY (US)-- has been inserted.

Signed and Sealed this

Seventeenth Day of October, 2006

JON W. DUDAS

Director of the United States Patent and Trademark Office